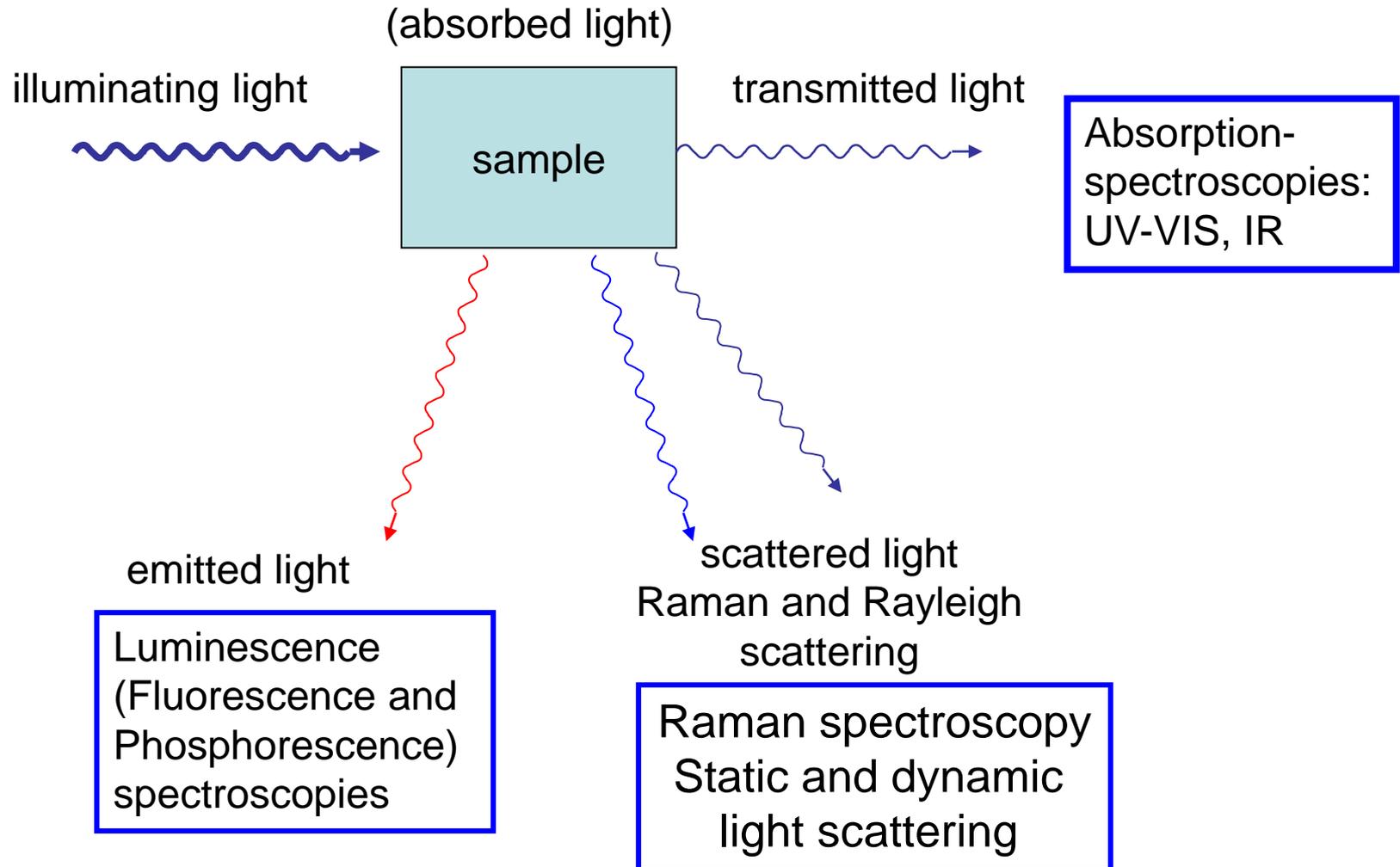


# Methods for investigation of macromolecular structures: IR, CD

# What happens if a sample is illuminated by light?



# Spectroscopy

(Absorption and emission spectroscopy)

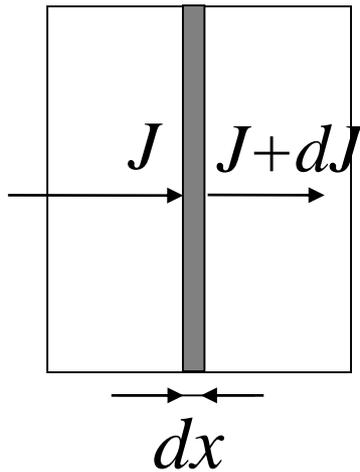
- Analysis of the wavelength dependence of the transmitted or emitted light.
- Information:
  - identification of atoms and molecules,
  - detection of changes in the molecular structure (conformation)
  - determination of the concentration





# Absorption spectroscopy

## The law of absorption



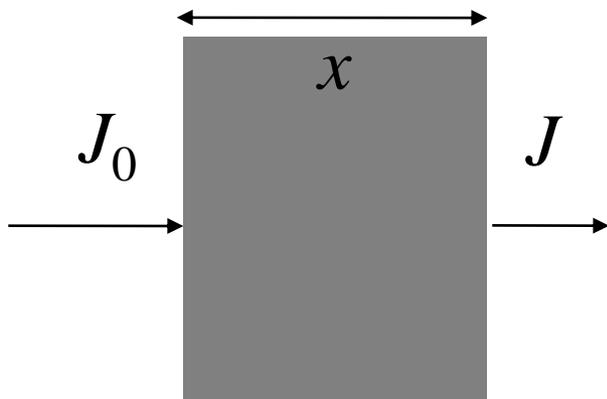
$$\left. \begin{array}{l} dJ \propto J \\ dJ \propto dx \end{array} \right\} dJ = -\mu J dx$$

$$\frac{dJ}{J} = -\mu dx$$

$$\int \frac{dJ}{J} = \int -\mu dx$$

$$\ln J = -\mu x + \text{const}$$

$$J = J_0 e^{-\mu x}$$



# Absorption spectroscopy

## Lambert-Beer law

Law of absorption:  $J=J_0 \cdot e^{-\mu x}$  where  $\mu(\text{material}, c, \lambda)$

Lambert-Beer law: 
$$A = \lg \frac{J_0}{J} = \varepsilon(\lambda)cx$$

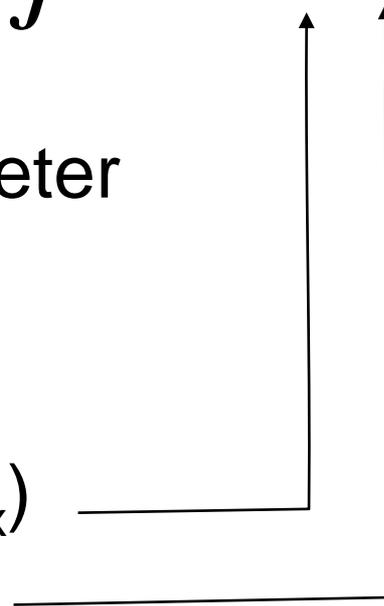
spectrum:  $A(\lambda)$

measurement: spectrophotometer

reference solution ( $J_0$ )

information: identification ( $\lambda_{\max}$ )

concentration (A)



# UV-VIS absorption spectroscopy

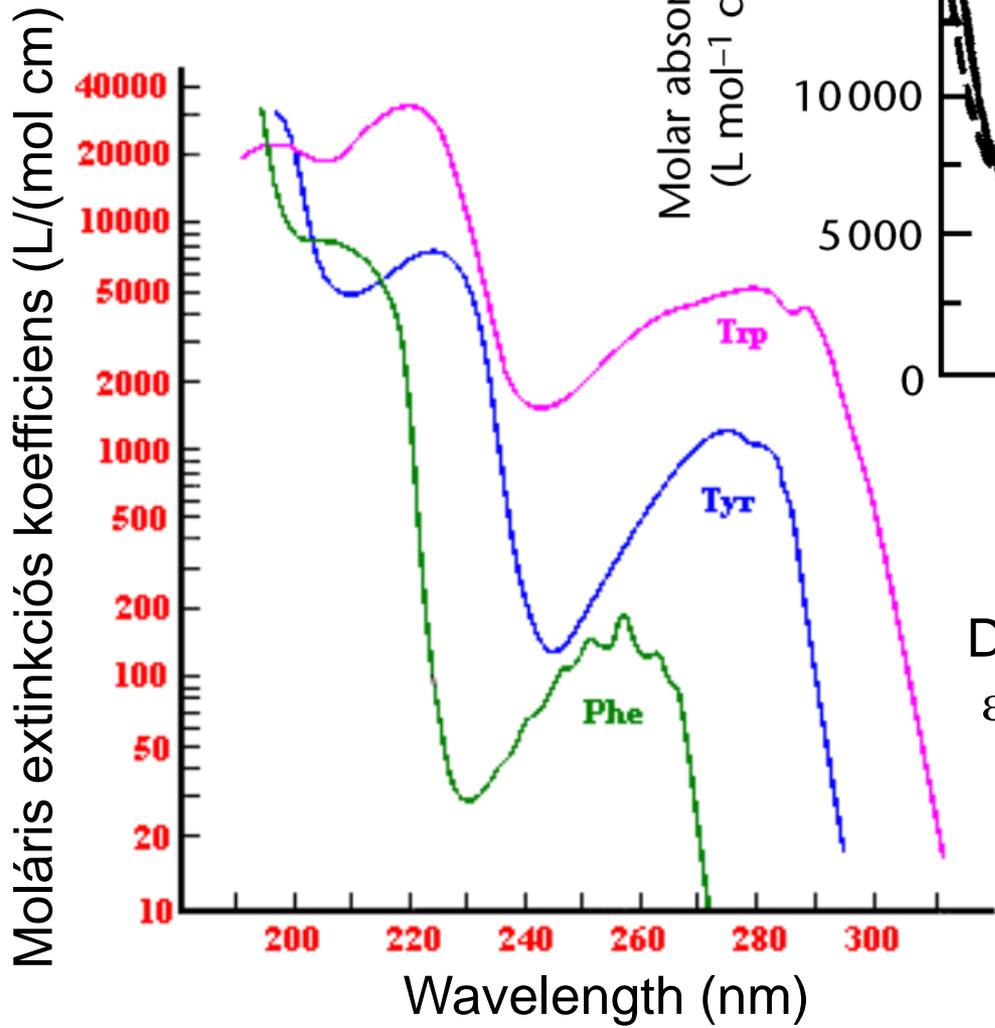
Proteins:

Absorbing site	$\lambda_{\max}$ (nm)	$\epsilon$ (L/cm mol)
Trp	280	5600
Tyr	274	1400
Phe	257	200
Disulfide bridge	250-270	300
Peptide bond	190-230	

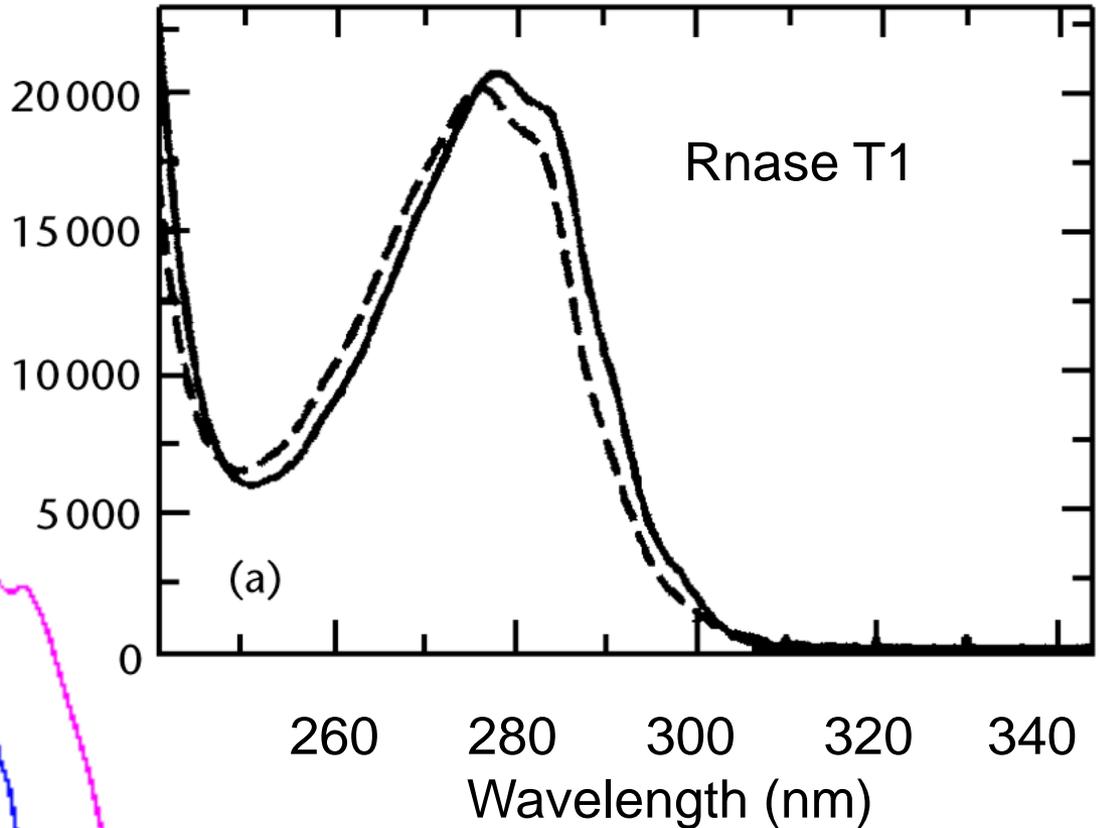
Determination of the protein concentration:

$$\epsilon_{280}(\text{L mol}^{-1}\text{cm}^{-1})=5500 n_{\text{Trp}} + 1490 n_{\text{Tyr}} + 125 n_{\text{SS}}$$

(Pace et al 1995. Prot. Sci. 4, 2411-242)



Molar absorbance  
(L mol<sup>-1</sup> cm<sup>-1</sup>)



Determination of the protein conc.:

$$\epsilon_{280}(\text{L mol}^{-1} \text{ cm}^{-1}) = 5500 n_{\text{Trp}} + 1490 n_{\text{Tyr}} + 125 n_{\text{SS}}$$

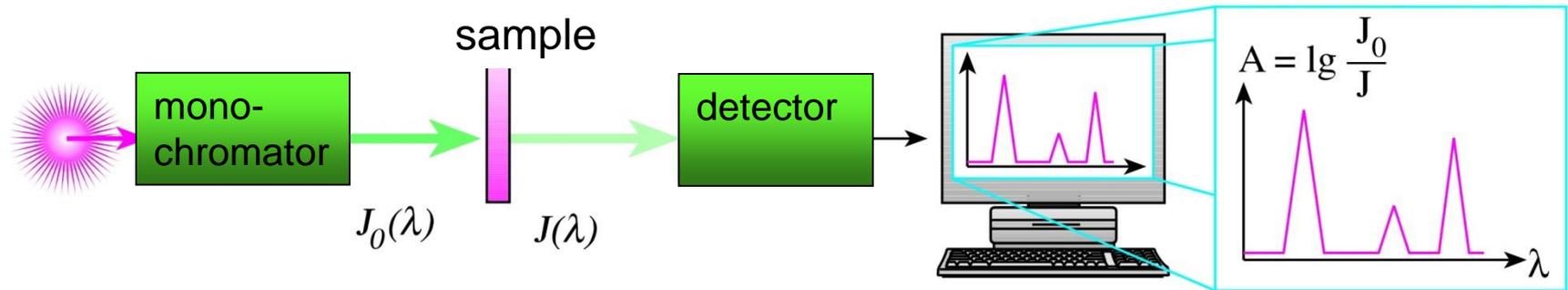
(Pace et al 1995. Prot. Sci. 4, 2411-242)

# Infrared spectroscopy

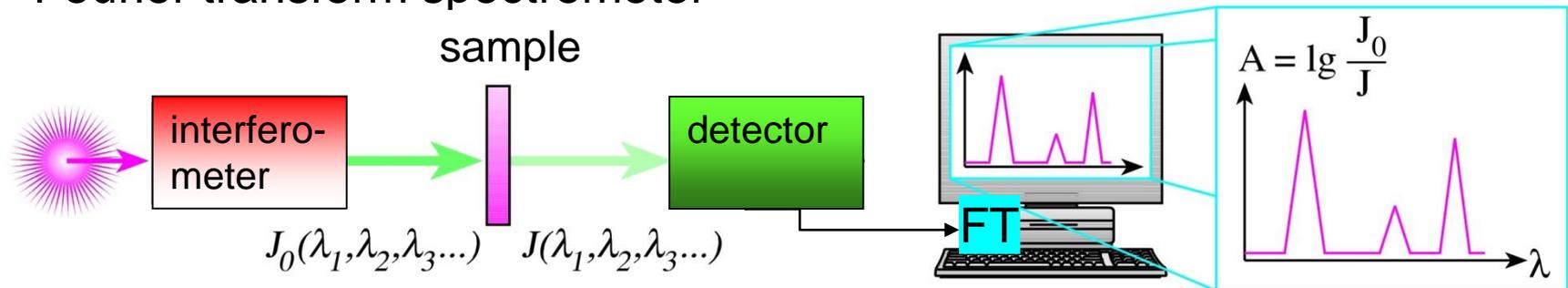
- Infrared light:  $\lambda=800 \text{ nm} - 1 \text{ mm}$   
MIR (mid-infrared) :  $2,5-50 \mu\text{m}$
- absorption spectroscopy
- the absorbed infrared radiation excite molecular vibrations
- very specific for the structure of the molecule
- special method for detection:  
FT spectrometer

# The technique of the measurement : Fourier transform spectrometer (FTIR)

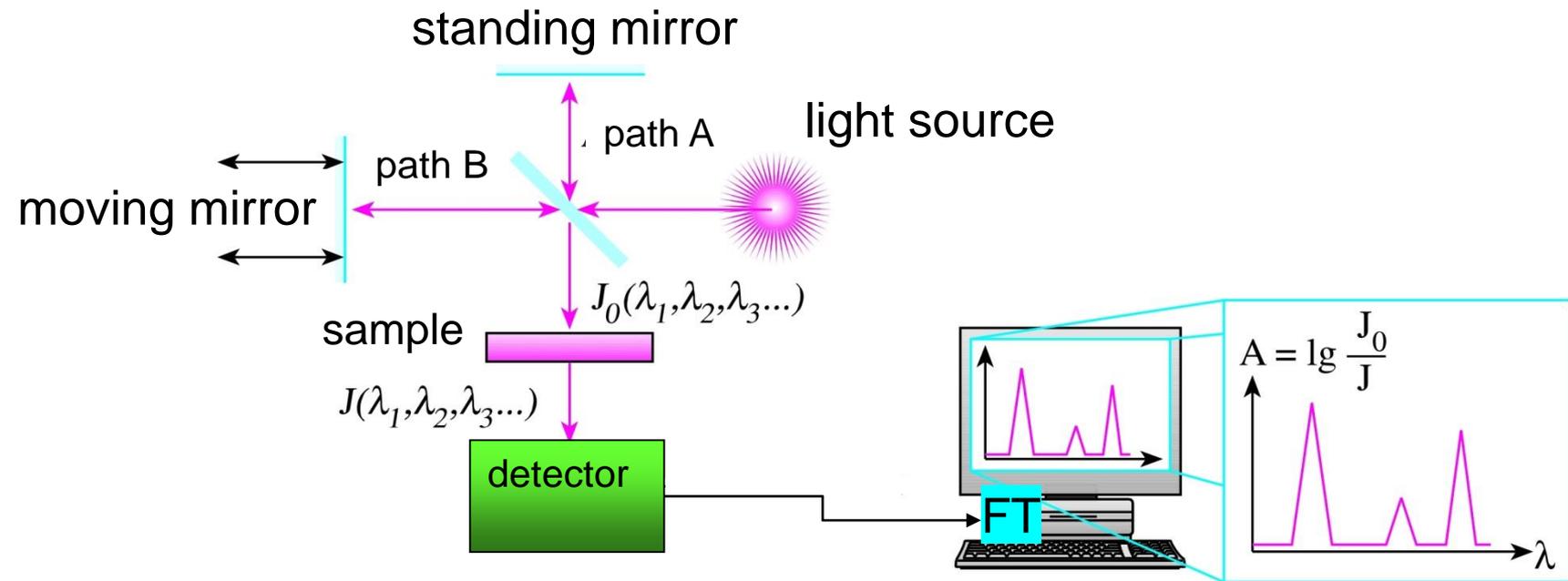
conventional (dispersion) spectrometer



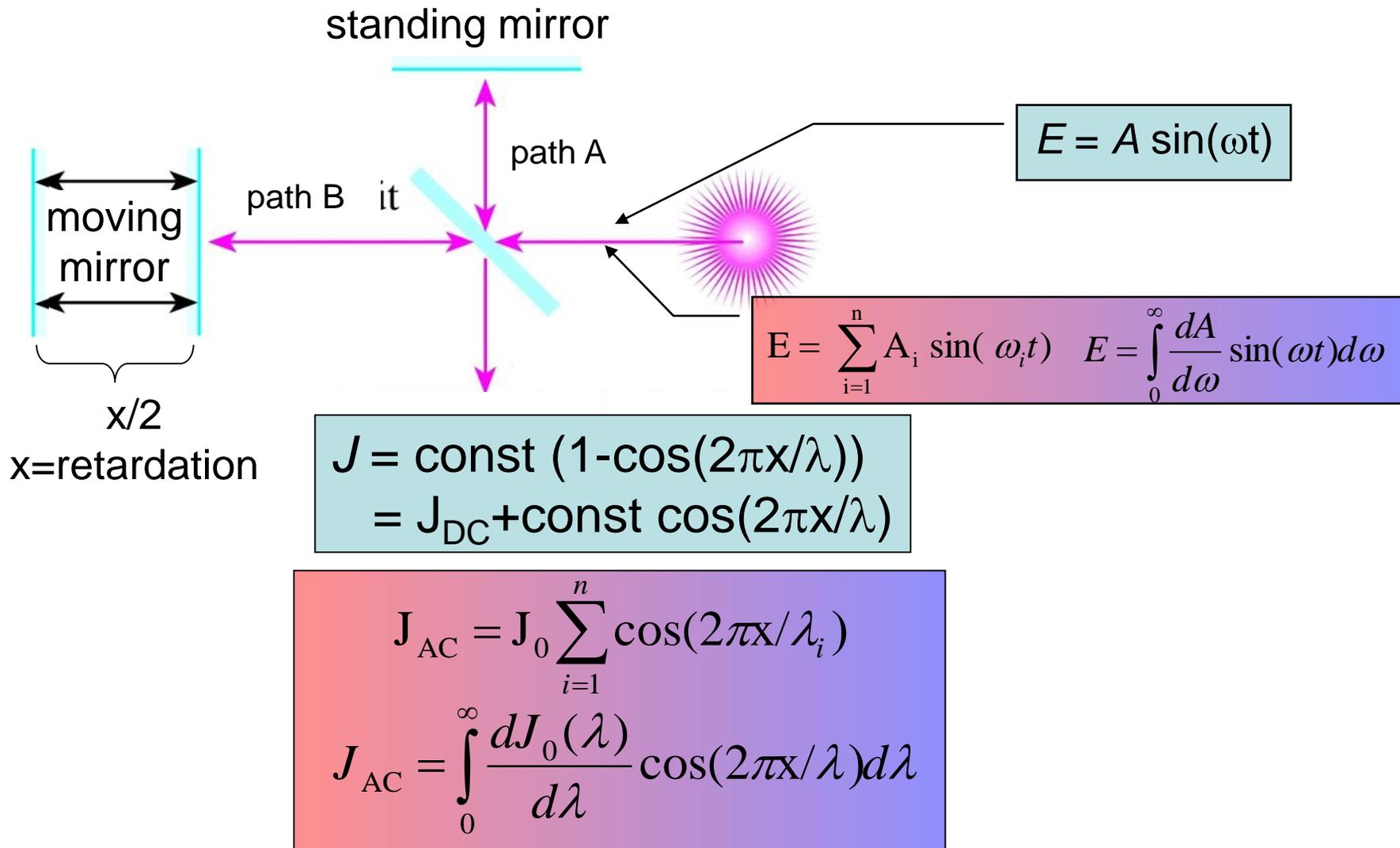
Fourier transform spectrometer



# The Fourier transform spectrometer



# Principle of the FTIR technique



# Fourier transformation

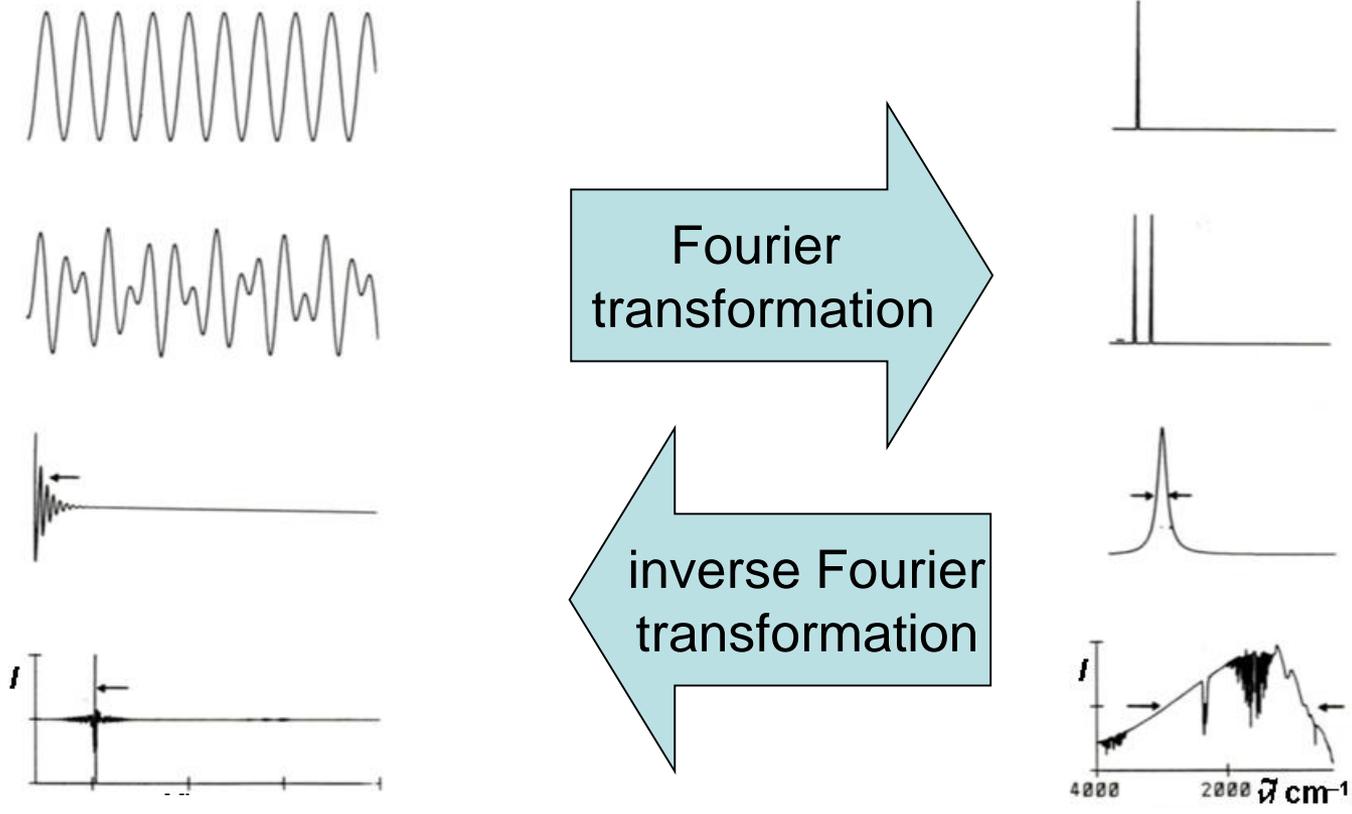
$g(\nu)$  is the Fourier transform of the function  $f(t)$ :

$$F(f(t)) = \int_{-\infty}^{\infty} f(t)e^{-2\pi i \nu t} dt = g(\nu)$$

Inverse Fourier transformation:

$$F^{-1}(g(\nu)) = \int_{-\infty}^{\infty} g(x)e^{2\pi i \nu t} d\nu = f(t)$$

# Fourier transformation



# How to obtain the spectrum in an FT spectrometer?

The intensity of the radiation passed through the interferometer:

$$J_{AC} = \int_0^{\infty} \frac{dJ_0(\lambda)}{d\lambda} \cos(2\pi x/\lambda) d\lambda$$

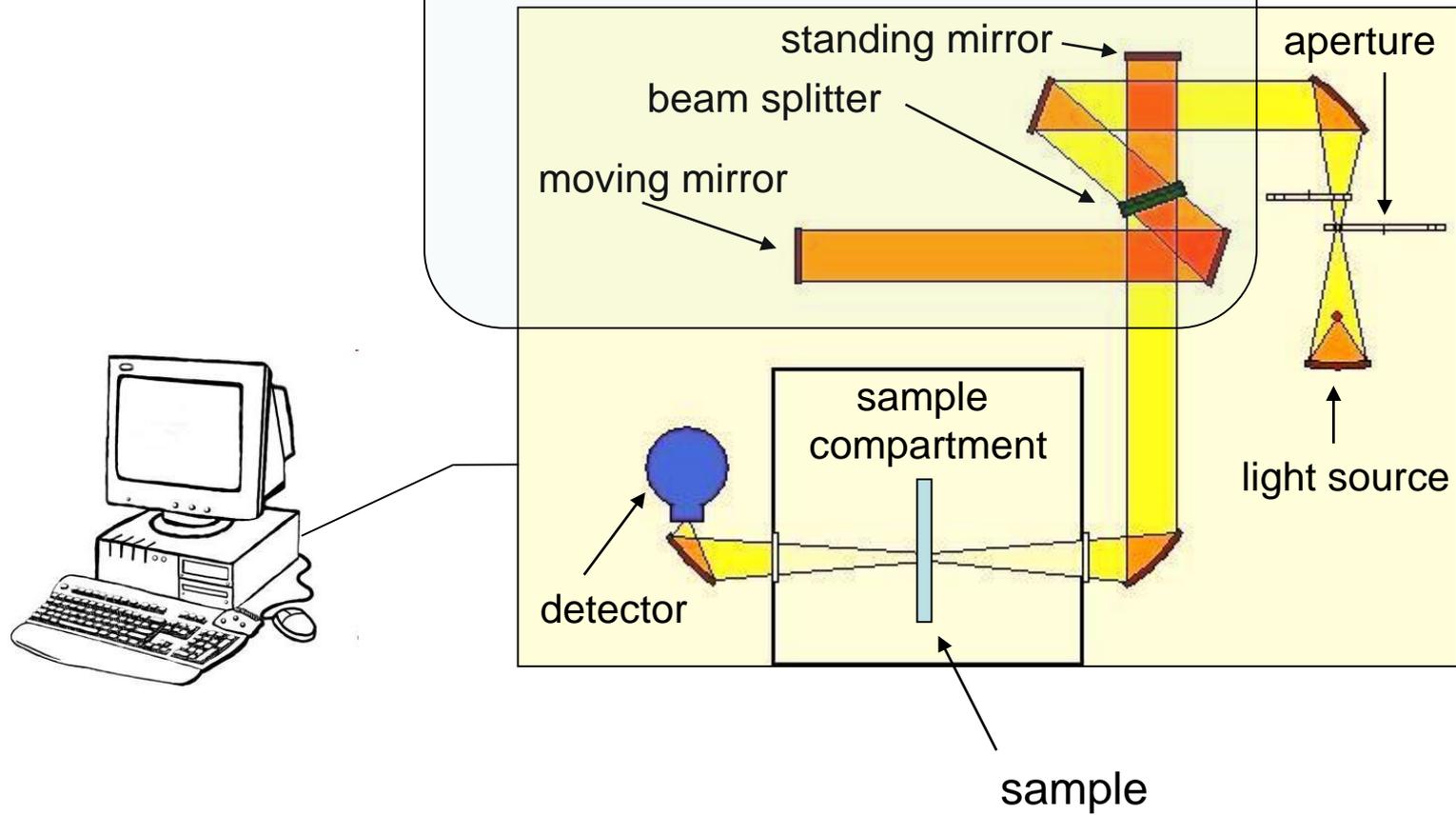
is the Cosine transform of  $\frac{dJ_0(\lambda)}{d\lambda}$

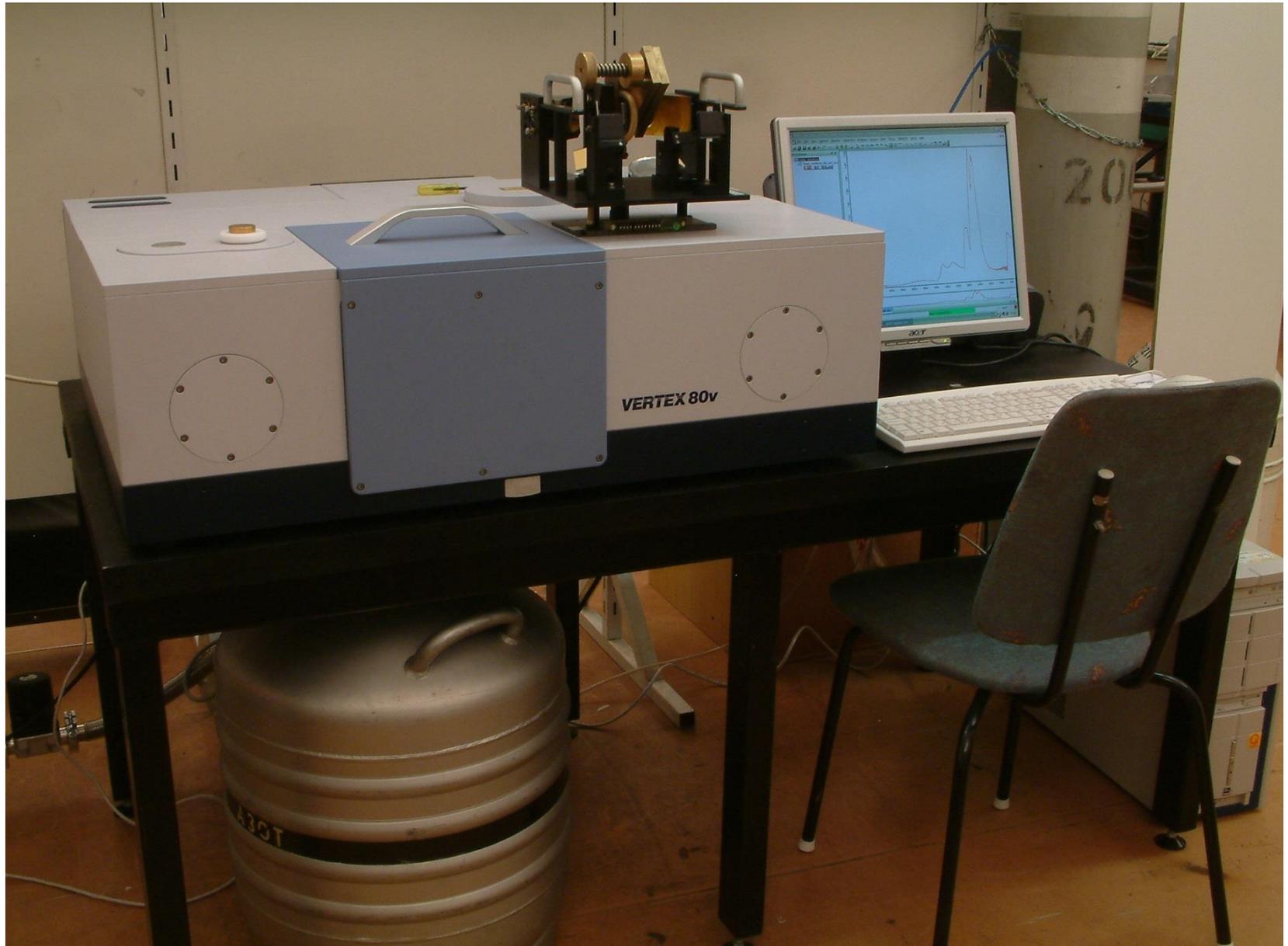
After the sample it reduces to  $\frac{dJ(\lambda)}{d\lambda}$

The transmission spectrum is:  $T(\lambda) = \frac{J(\lambda)}{J_0(\lambda)} = \frac{\frac{dJ(\lambda)}{d\lambda}}{\frac{dJ_0(\lambda)}{d\lambda}}$

The absorption spectrum is:  $A(\lambda) = \lg (1/T(\lambda))$

# Michelson interferometer



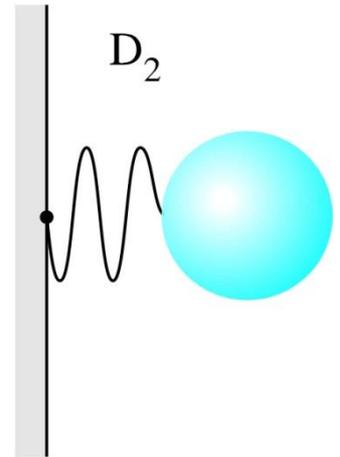
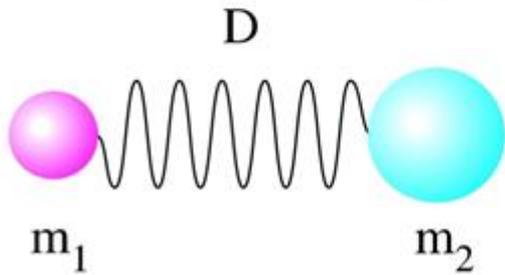
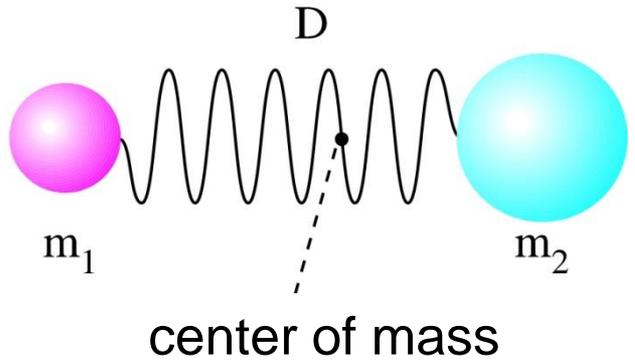
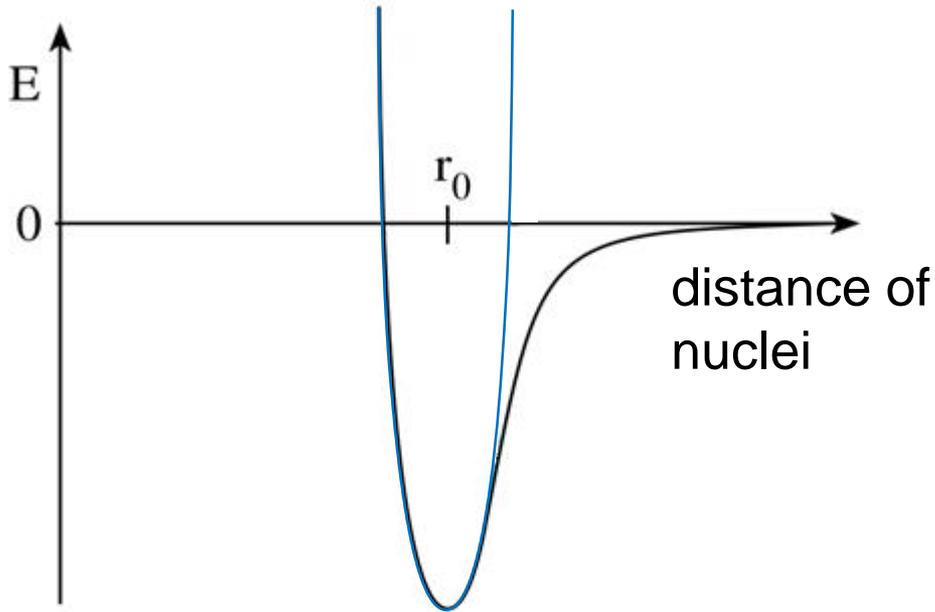


# Molecular vibrations

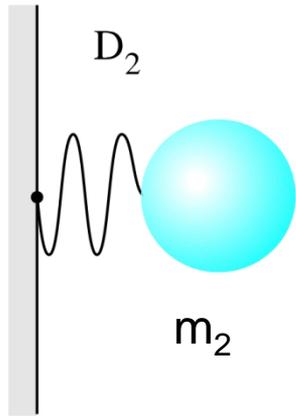
The electrons are light ( $m_e \ll m_{\text{nucleus}}$ ), they can follow the movements of the nuclei easily, therefore the movements of the nuclei are independent of the movements of the electrons.

Classical physical description: the chemical bond is represented by a spring.

# Molecular vibrations:

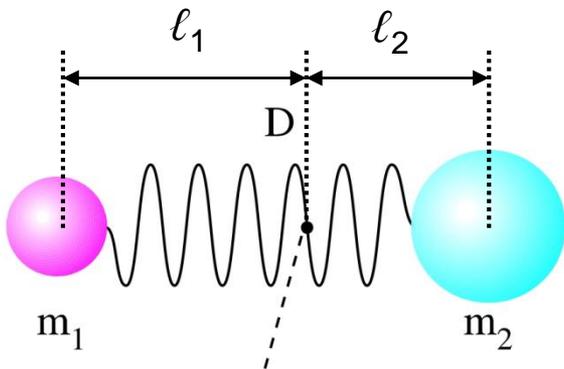


known from elementary mechanics:



$$f = \frac{1}{2\pi} \sqrt{\frac{D_2}{m_2}}$$

$$\frac{m_2}{m_1} = \frac{l_1}{l_2}$$

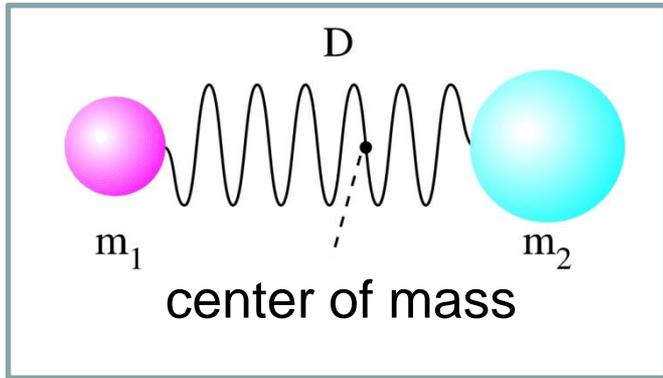


center of mass

$$F = D\Delta l$$

$$\begin{aligned} \frac{D_2}{D} &= \frac{F/D}{F/D_2} = \frac{\Delta l}{\Delta l_2} = \frac{l}{l_2} = \\ &= \frac{l_1 + l_2}{l_2} = \frac{l_1}{l_2} + 1 = \frac{m_2}{m_1} + 1 = \frac{m_1 + m_2}{m_1} \end{aligned}$$

substituting  $\frac{m_1 + m_2}{m_1} = \frac{D_2}{D}$  into  $f = \frac{1}{2\pi} \sqrt{\frac{D_2}{m_2}}$



The frequency of the vibration:

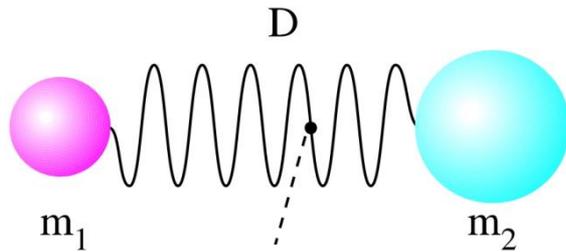
$$f = \frac{1}{2\pi} \sqrt{\frac{D(m_1 + m_2)}{m_1 m_2}}$$

$$m_{red} = \frac{m_1 m_2}{m_1 + m_2}$$

is called as reduced mass

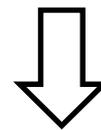
Frequency with the reduced mass:  $f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$

# How can we induce molecular vibrations?



$$f_{E.M. rad} = f_{vibration} = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$

Resonance



absorption of the  
energy of the radiation

The wavelength of this radiation is:

$$\lambda = \frac{c}{f} = 2\pi c \sqrt{\frac{m_{red}}{D}}$$

In the IR spectroscopy the wavenumber ( $\nu$ ) is used, which is the reciprocal of  $\lambda$ :

$$\nu = \frac{1}{\lambda} = \frac{1}{2\pi c} \sqrt{\frac{D}{m_{red}}}$$

$\nu$ : number of waves in a unit length [ $\text{cm}^{-1}$ ]

Example: CO

The measured wavenumber:  $\nu = 2143 \text{ cm}^{-1}$

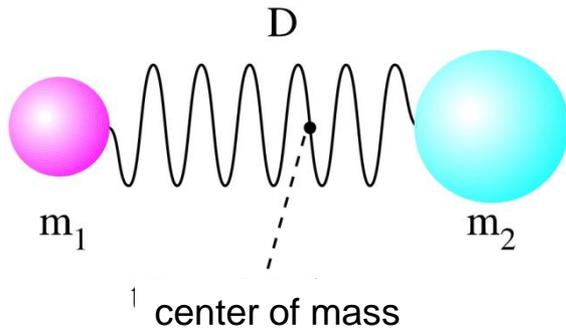
$$\left. \begin{array}{l} \Rightarrow \lambda = 4,67 \mu\text{m} \Rightarrow f = 6,43 \cdot 10^{13} \text{ Hz} \\ m_{\text{C}} = 2 \cdot 10^{-26} \text{ kg}, \quad m_{\text{O}} = 2,7 \cdot 10^{-26} \text{ kg} \end{array} \right\} \Rightarrow D = 1875 \text{ N/m}$$

if  $\nu$  is known,  $D$  can be calculated

if  $D$  is known,  $\nu$  can be calculated

# Classical vs. quantum physics

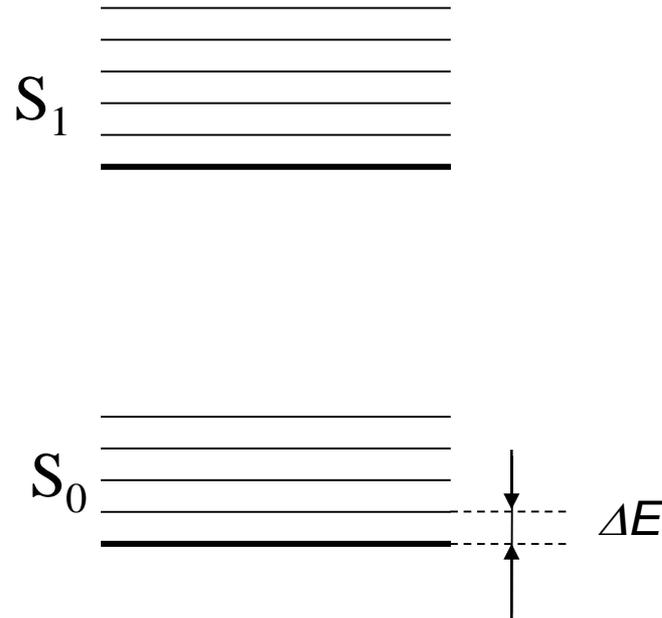
Classical physical picture



$$f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$

resonance with the light with frequency  $f$

Quantum mechanical picture



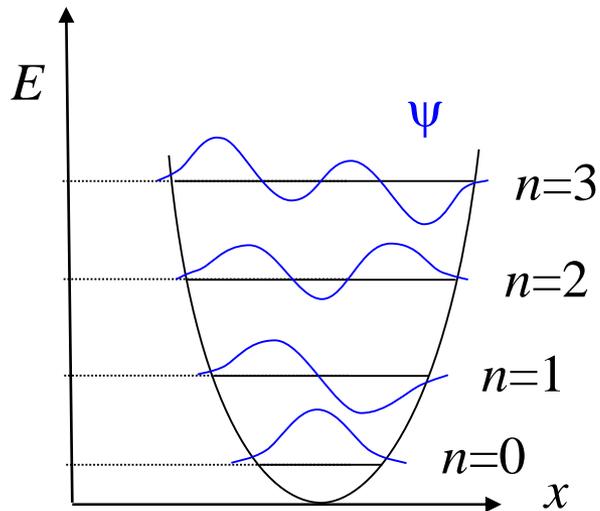
$$\Delta E = hf$$

A yellow box containing an equals sign (=) is positioned between two arrows. One arrow points from the box to the frequency  $f$  in the equation above, and the other points from the box to the energy difference  $\Delta E$  in the equation above.

# Quantummechanical description

Quantum harmonic oscillator

Particle in a harmonic potential



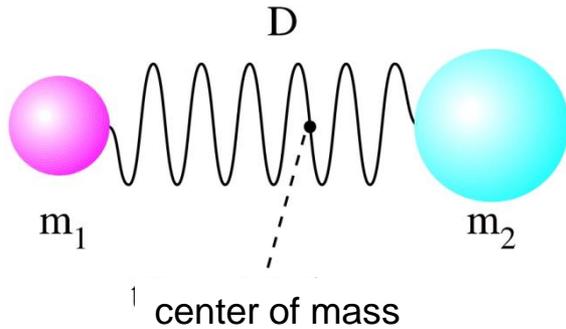
Energy eigenvalues

$$E_n = hf \left( n + \frac{1}{2} \right)$$

$$n = 0, 1, 2, \dots$$

# Classical vs. quantum physics

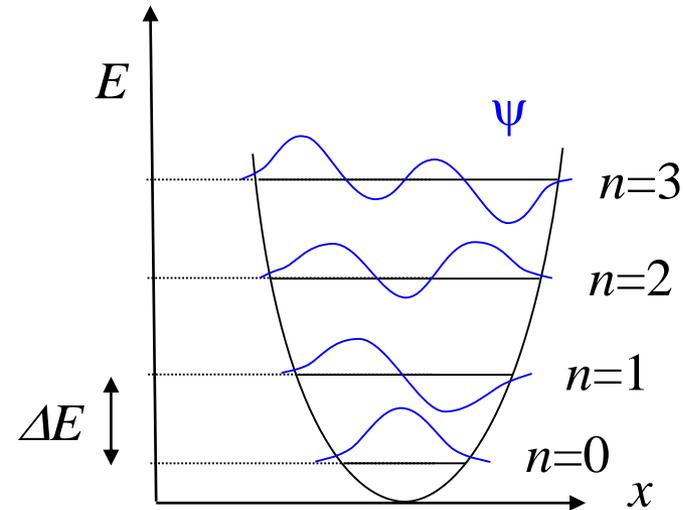
Classical physical picture



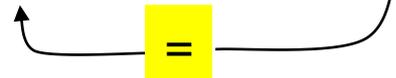
$$f = \frac{1}{2\pi} \sqrt{\frac{D}{m_{red}}}$$

resonance with the light with frequency  $f$

Quantum mechanical picture



$$\Delta E = hf$$



# Dependence of the vibrational frequency on the mass of the atoms and on the strength of the chemical bond.

mass: 

## Infravörös rezgési frekvenciák (cm<sup>-1</sup>)

<b>B-H</b> 2400	<b>C-H</b> 3000	<b>N-H</b> 3400	<b>O-H</b> 3600	<b>F-H</b> 4000
<b>Al-H</b> 1750	<b>Si-H</b> 2150	<b>P-H</b> 2350	<b>S-H</b> 2570	<b>Cl-H</b> 2890
	<b>Ge-H</b> 2070	<b>As-H</b> 2150	<b>Se-H</b> 2300	<b>Br-H</b> 2650

Water (O-H): 3600 => D<sub>2</sub>O: 2600 cm<sup>-1</sup>

Bond strength:

C-N: 1100 cm<sup>-1</sup>,

C=N: 1660 cm<sup>-1</sup>,

C≡N: 2220 cm<sup>-1</sup>.

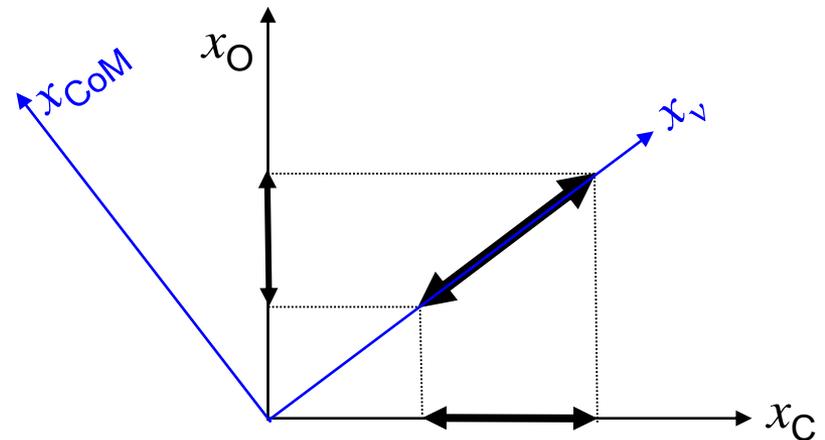
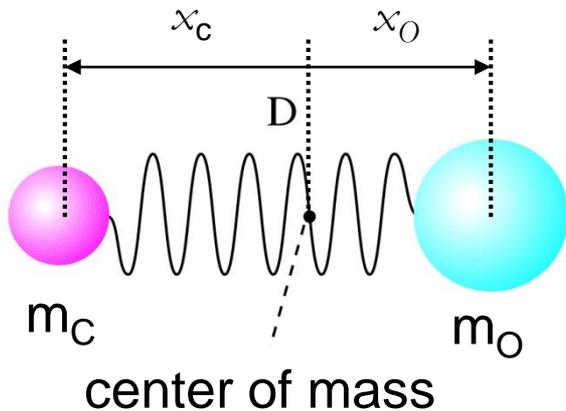
# Vibrations of the large molecules

Molecule consisting of  $N$  atoms:

- $3N$  degree of freedom,  
     $3-3$  are the rotations and translations  
    of the whole molecule
- $3N-6$  vibrational degree of freedom  
    ( $3N-5$  for the linear molecules)
- $3N-6$  independent normal vibrations

# Normal coordinates

Shown in case of a simple molecule of two atoms:



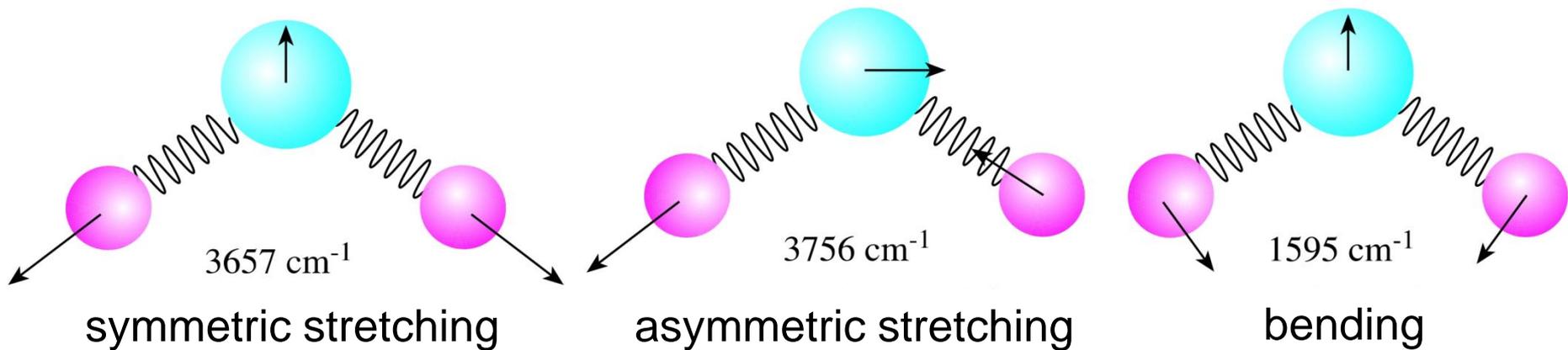
In general case one has to rotate a  $3N$  dimensional coordinate system.  
Linear transformation (matrix operation)

# Normal vibrations

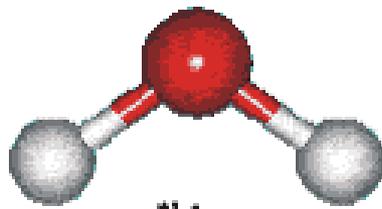
All the atoms vibrate

- with the same frequency but
- with different amplitude and
- in different direction.

Example: water

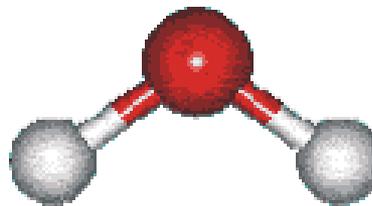


# Normal vibrations of water



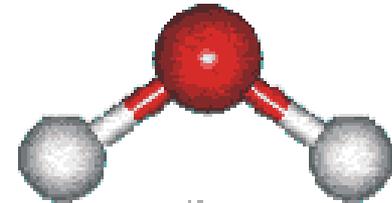
$\nu_1$

symmetric stretch



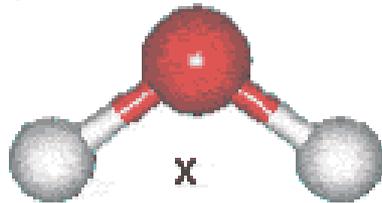
$\nu_3$

asymmetric stretch

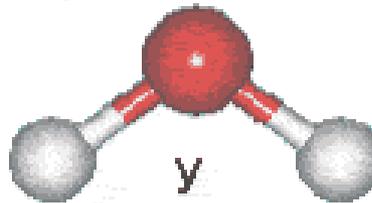


$\nu_2$

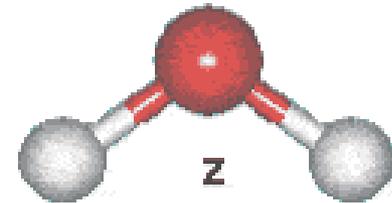
bend



x



y

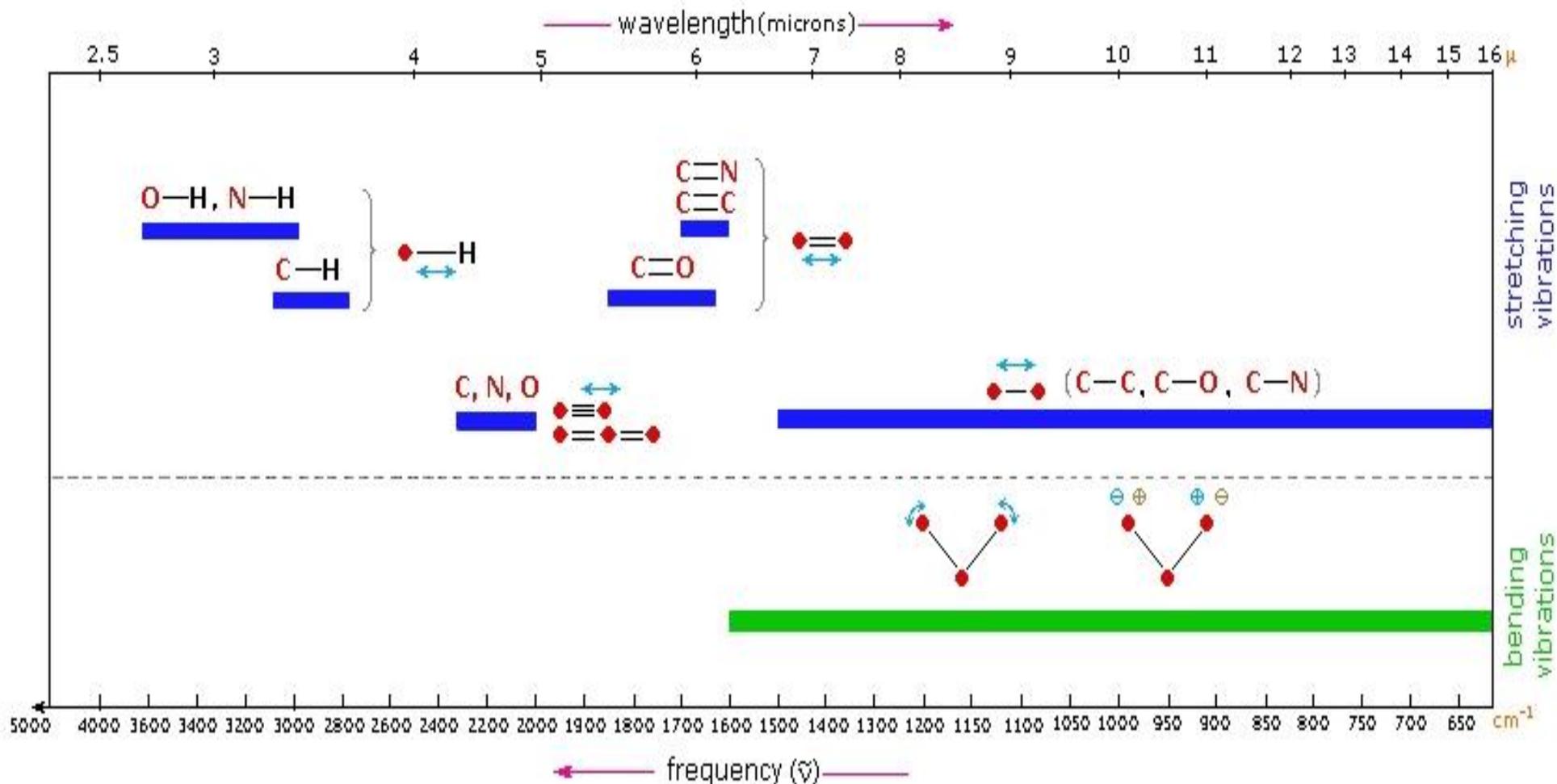


z

librations

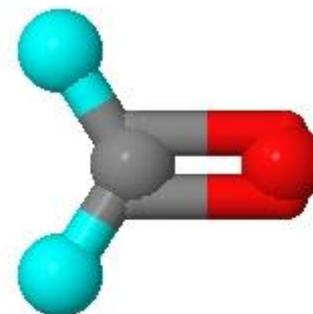
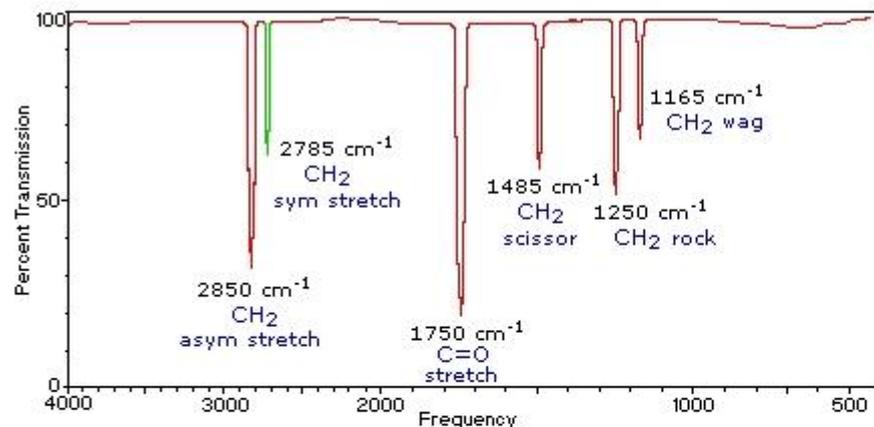
These are no vibrations! These are rotations!

# Typical vibrational frequencies (wavenumbers)



# Example: Formaldehyde

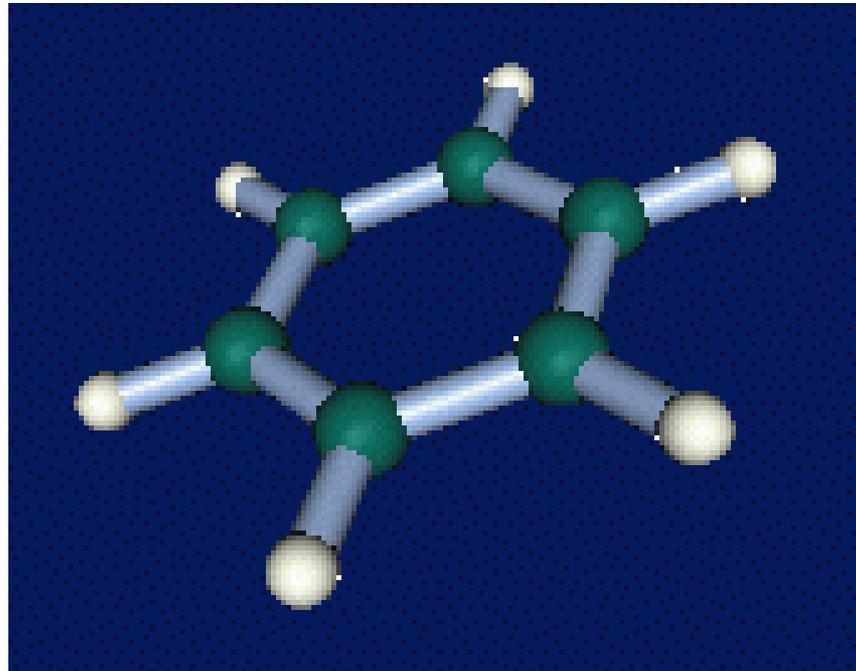
Gas Phase Infrared Spectrum of Formaldehyde,  $\text{H}_2\text{C}=\text{O}$

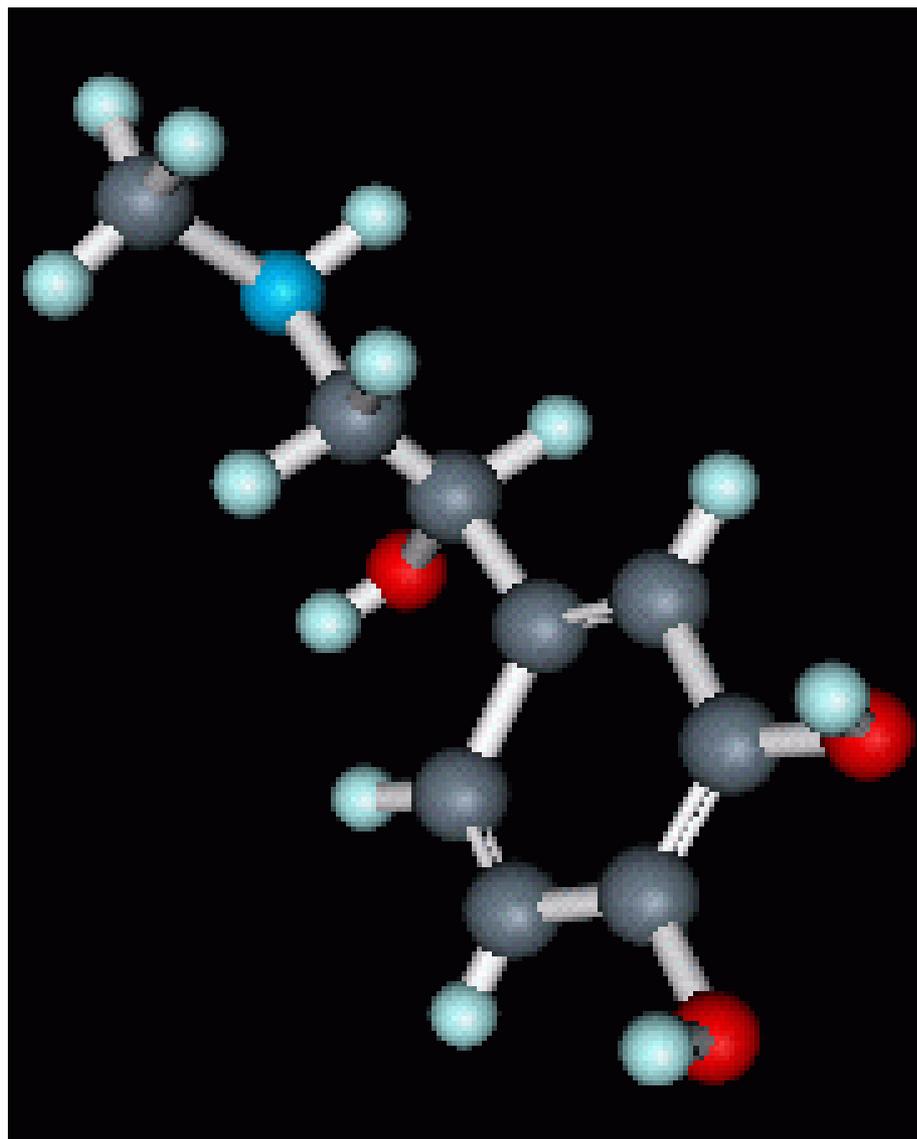


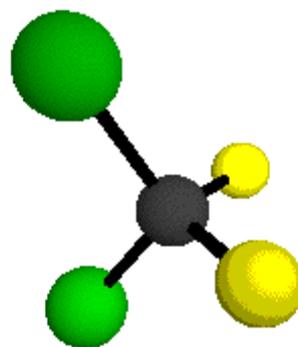
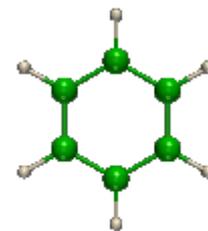
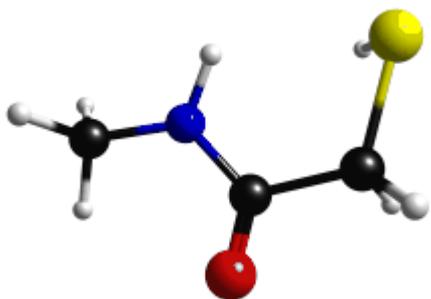
- View  $\text{CH}_2$  Asymmetric Stretch
- View  $\text{CH}_2$  Symmetric Stretch
- View  $\text{C}=\text{O}$  Stretch
- View  $\text{CH}_2$  Scissoring
- View  $\text{CH}_2$  Rocking
- View  $\text{CH}_2$  Wagging

- Ball&Stick Model
- Spacefill Model
- Stick Model
- Motion Off

# Benzol





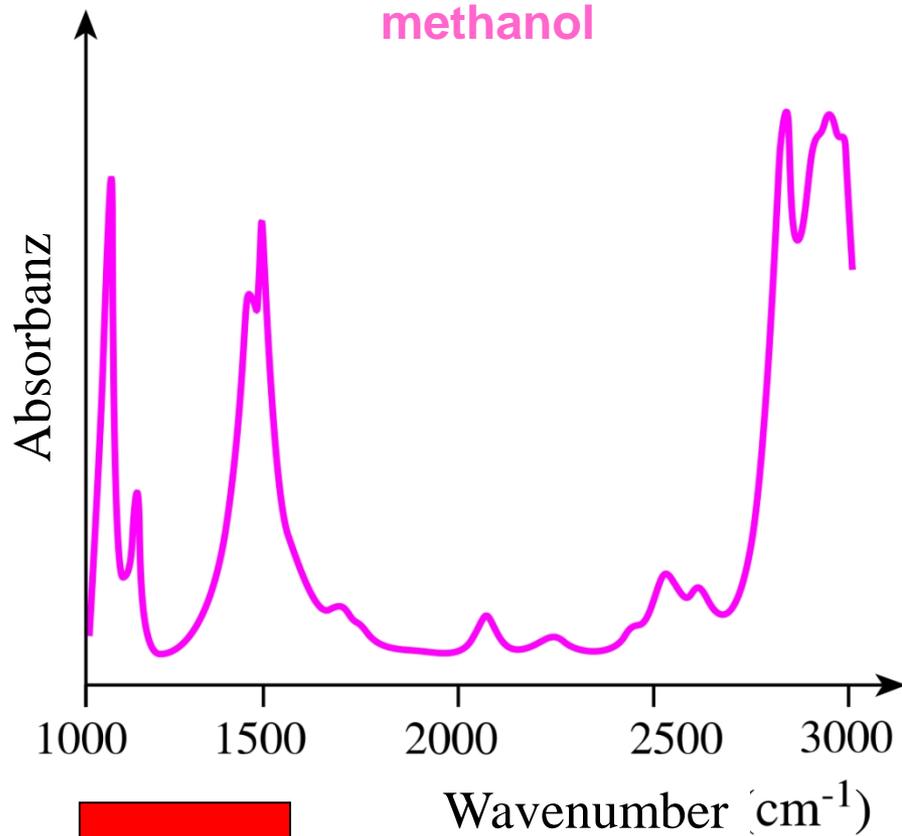


# Analytical applications

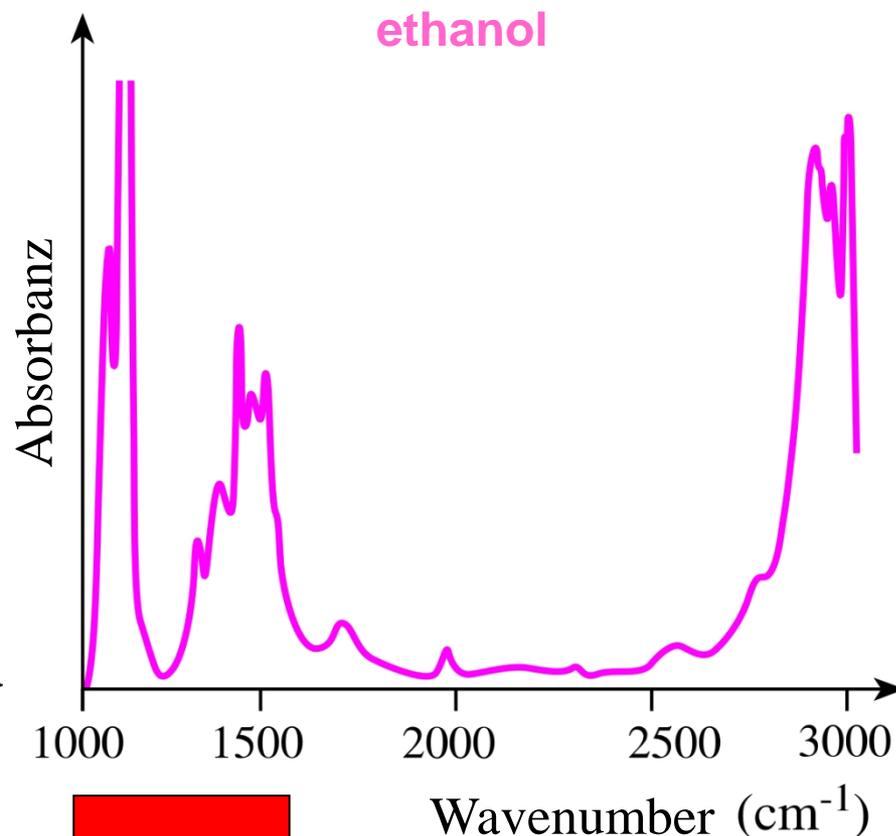
- synthesis: identification of the intermediate and the end product
- determination and justification of the molecular structure
- detection of the metabolites
- quality control (purity)
  
- Remark.: Lambert-Beer law is valid, determination of concentration is possible

# Identification of the Molecules

methanol



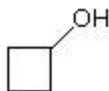
ethanol



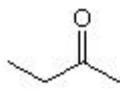
Fingerprint Region

# Example: Identification of molecules

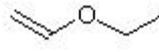
$C_4H_8O$



cyclobutanol



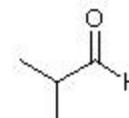
2-butanone



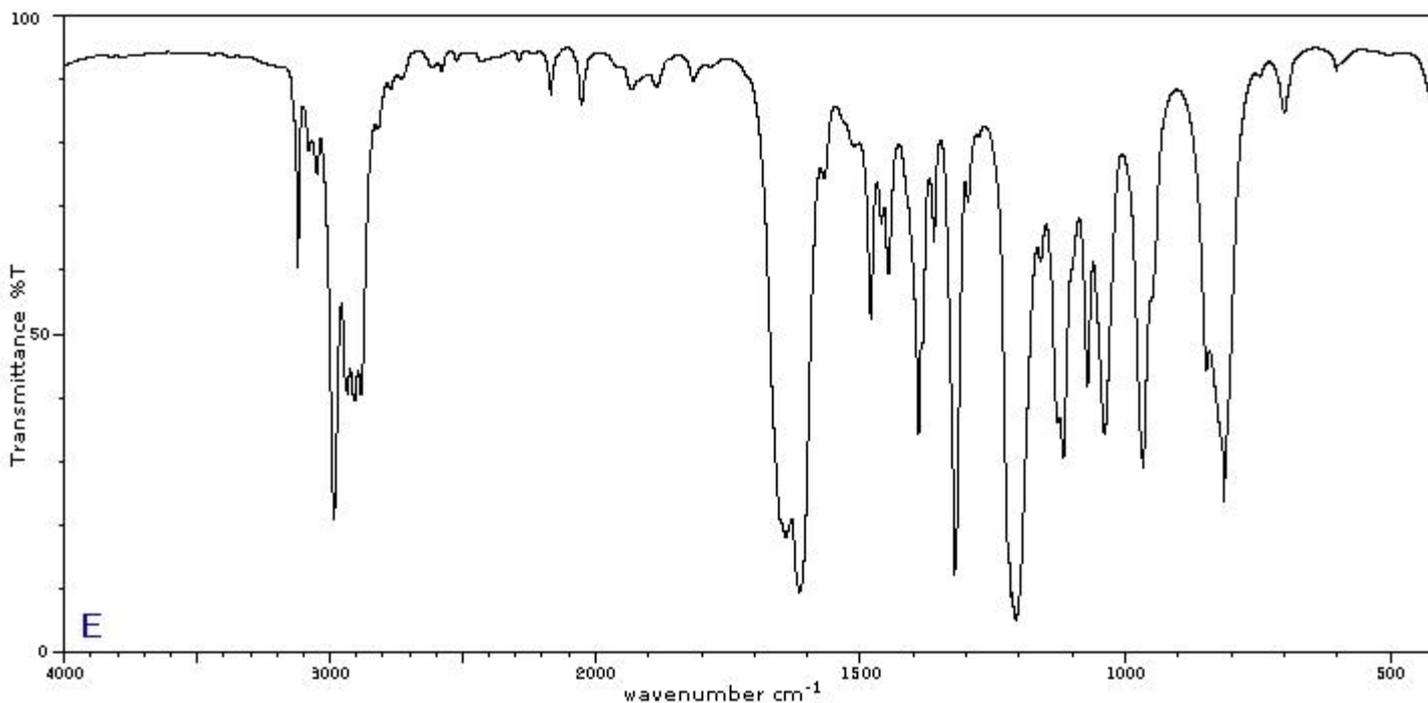
ethyl vinyl ether



2-methyl-2-propen-1-ol



2-methylpropanal



**Fingerprint Region**

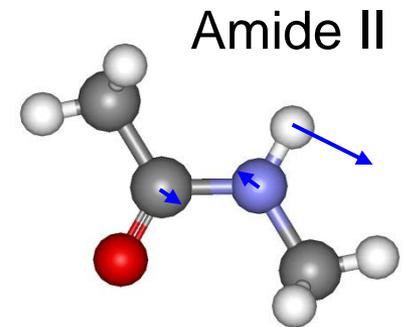
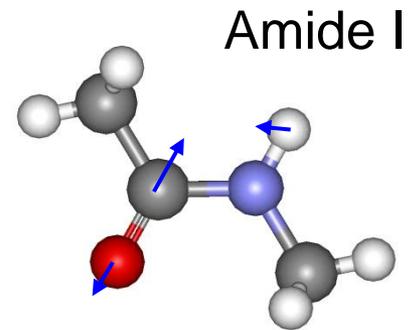
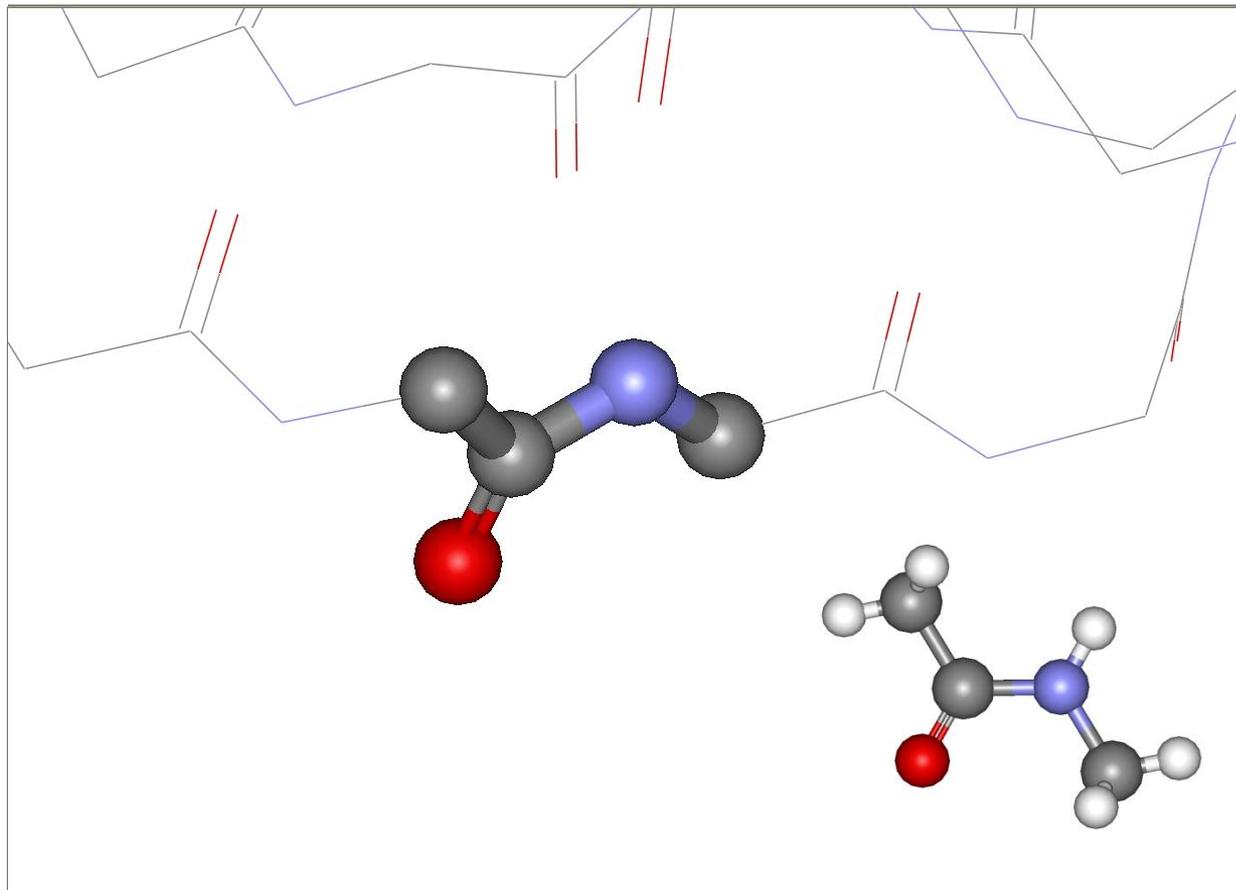
# Vibrations of the macromolecules

Complex global vibrations

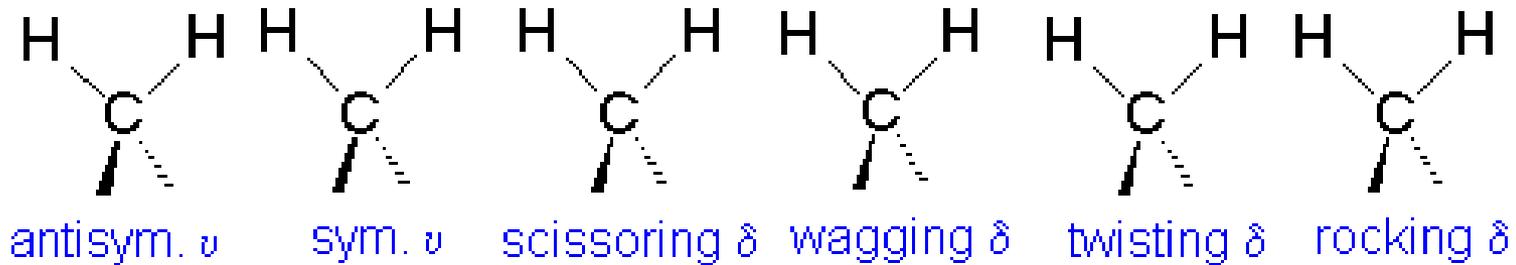
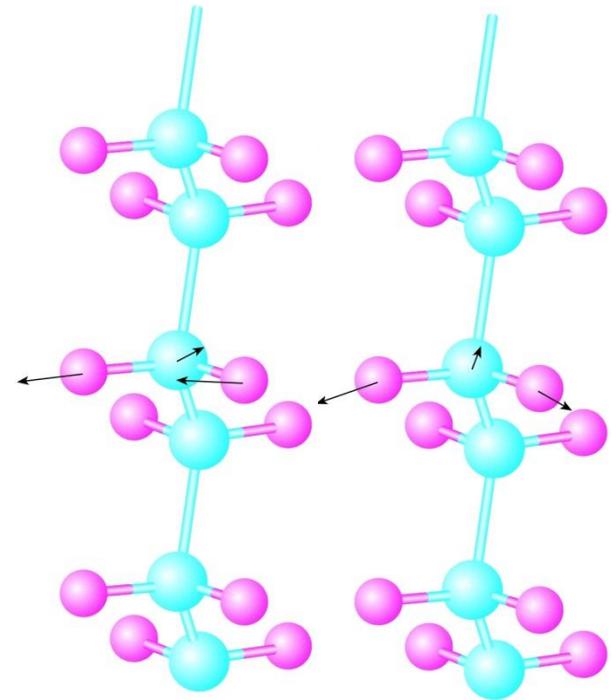
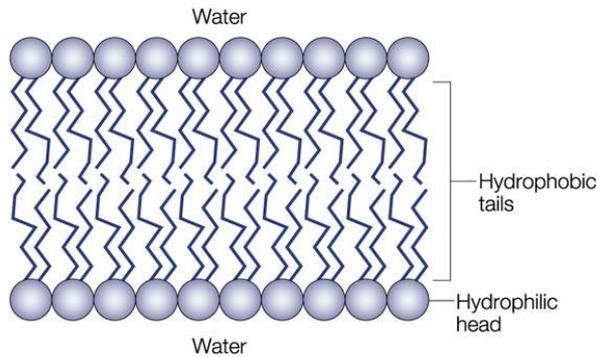
Localized vibrations, e.g.:

- amid vibrations of proteins  
(obtained from N-methylacetamide)
- CH<sub>2</sub> vibrations of the lipids

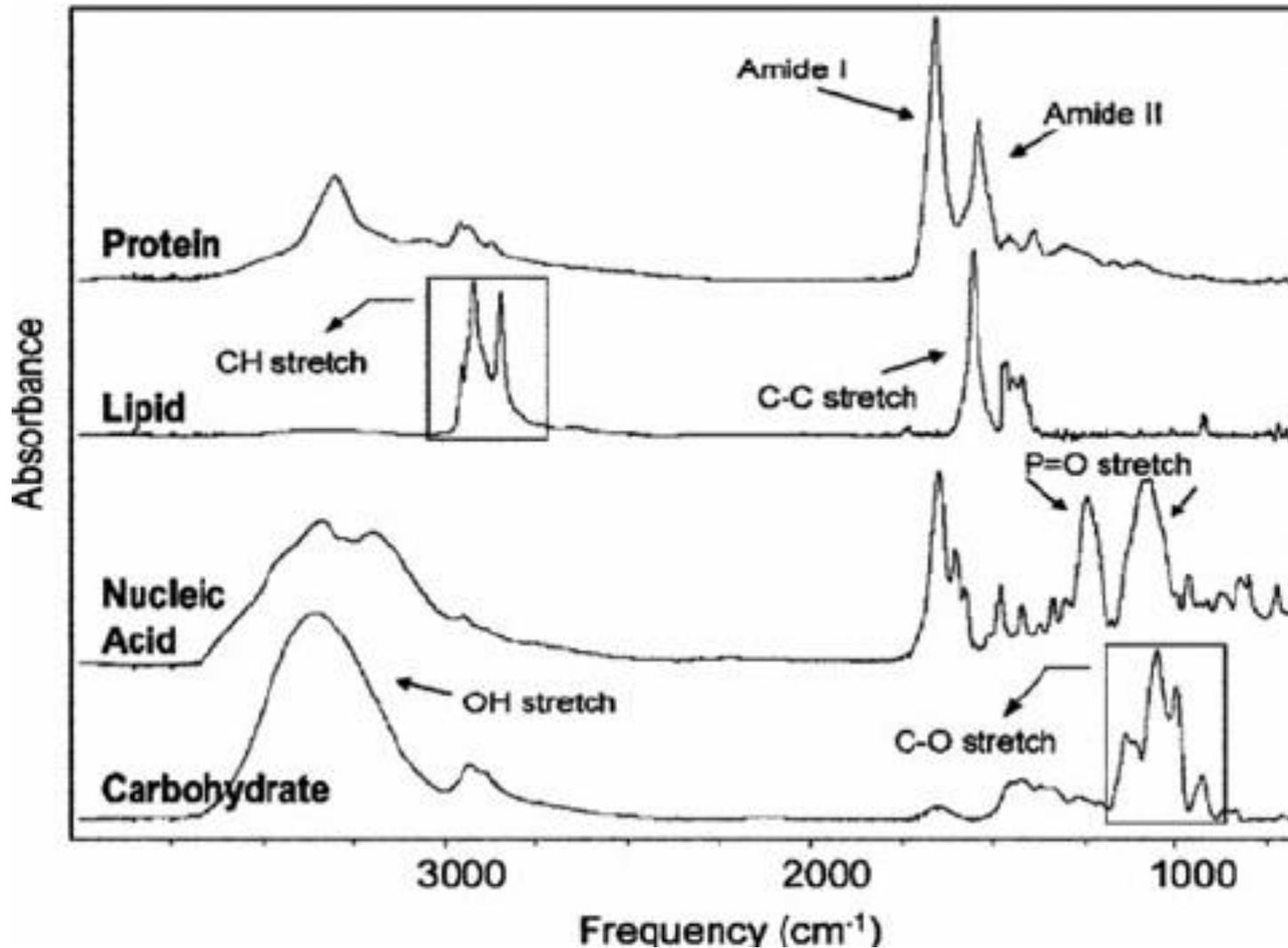
# Proteins: model of the polypeptide backbone



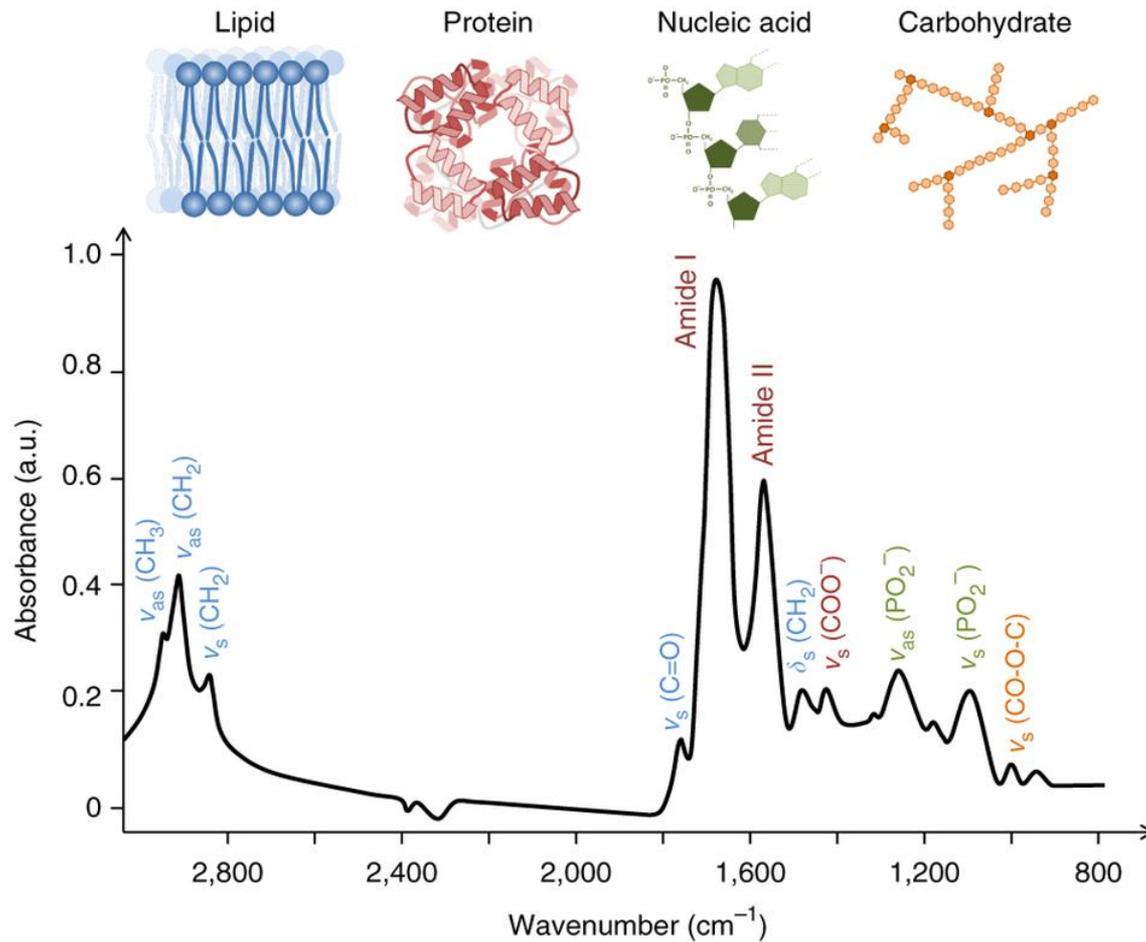
# Localized vibrations: CH<sub>2</sub> vibrations of the lipids



# Typical infrared spectra of biological macromolecules



# Infrared spectrum of the cell

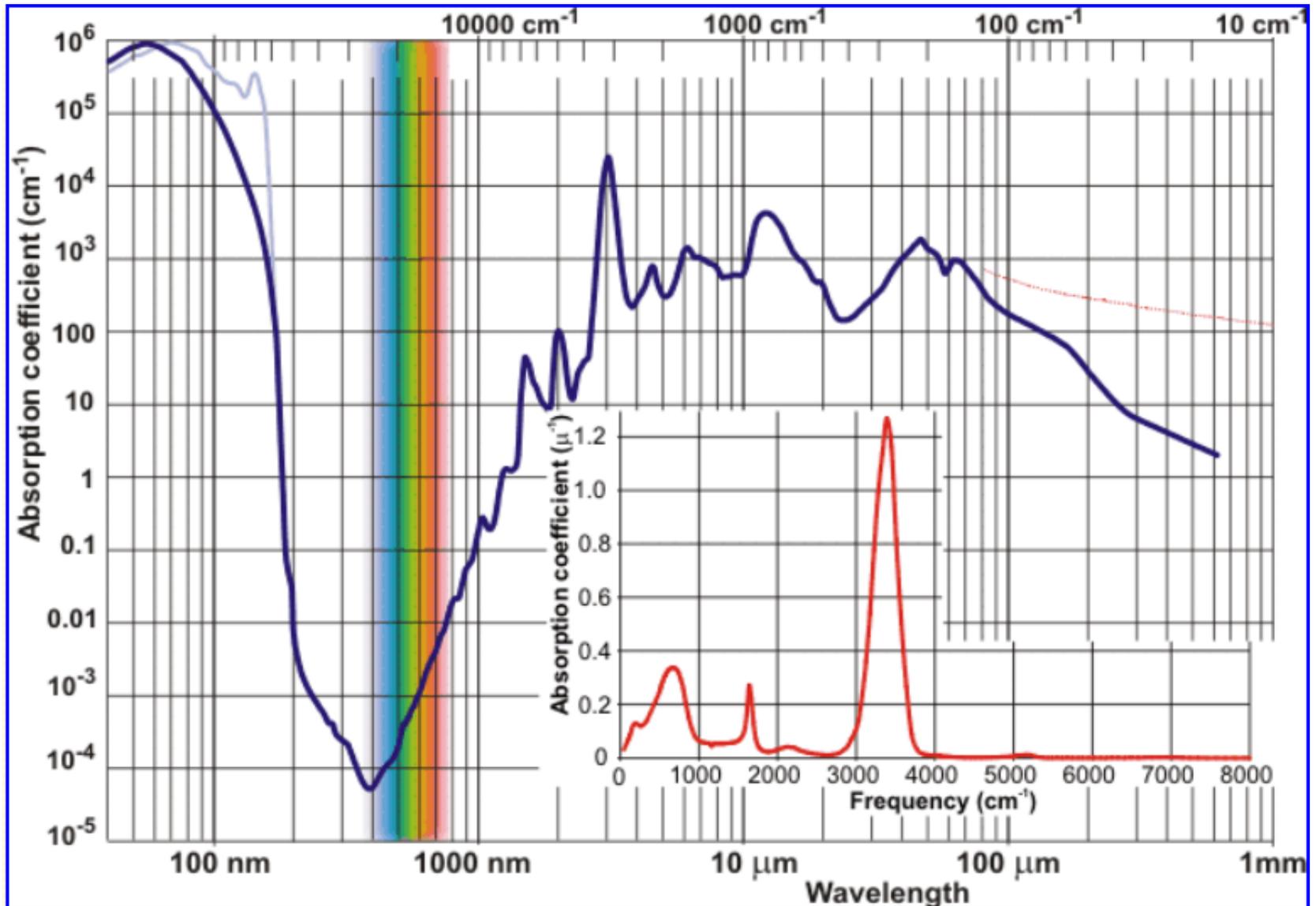


# IR spectroscopy of proteins

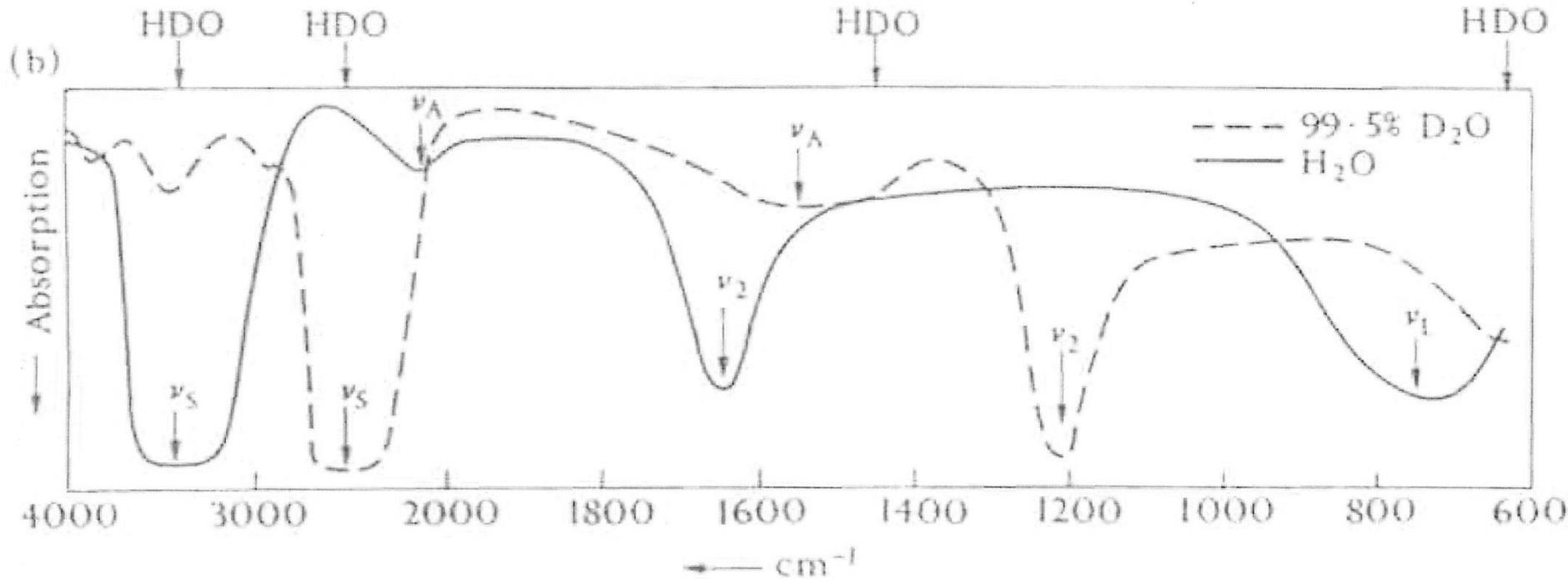
- Backbone: amide vibrations
  - conformation (secondary structure)
  - H/D exchange, (tertiary structure)
- Side chains
  - interaction with other molecules
  - $\text{Ca}^{2+}$  binding

Technical point: heavy water solutions ( $\text{D}_2\text{O}$ )

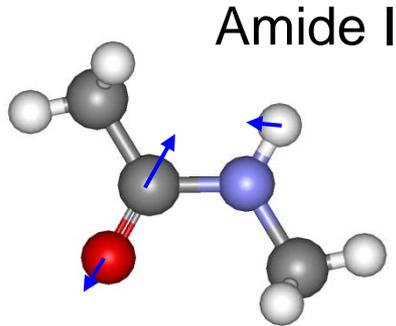
# Absorption spectrum of the water



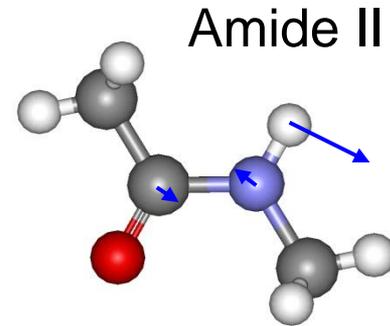
# Absorption spectra of water and heavy water



# Amide vibrations of proteins

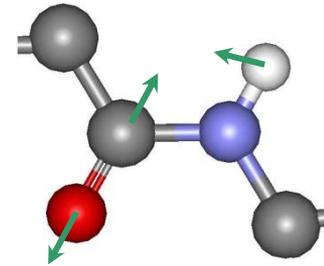


amide I  
C=O vibration.  
Sensitive to the  
secondary  
structure  
because of the  
hydrogen bond.

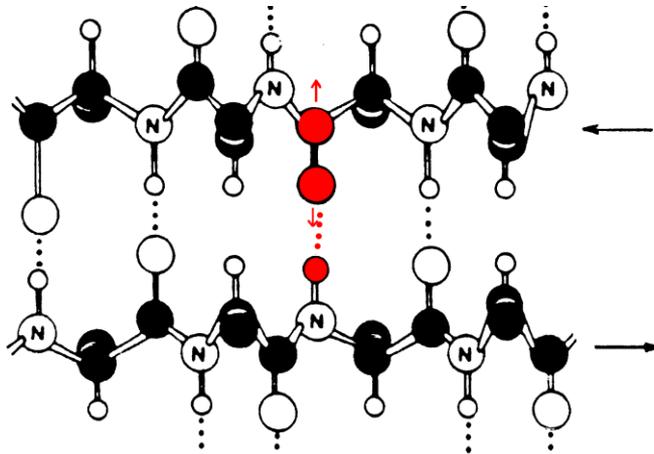


amide II  
N-H in plane bending  
vibration  
Sensitive to the H-D  
exchange i.e. to the  
compactness of the  
molecule.  
(tertiary structure)

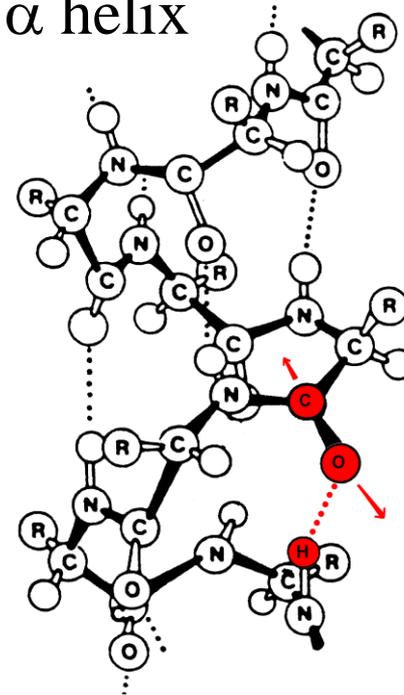
# The amide I vibration and the secondary structure



$\beta$  sheet



$\alpha$  helix



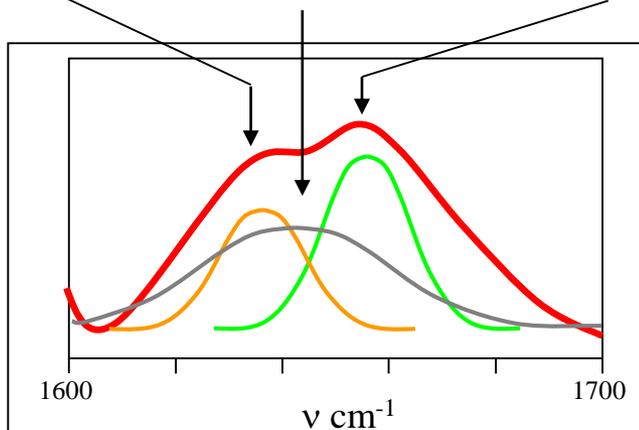
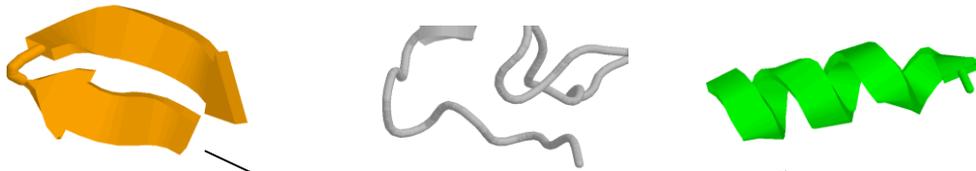
# Position of the amide I band in different secondary structures

## Intramolecular structure

$\beta$ -sheet

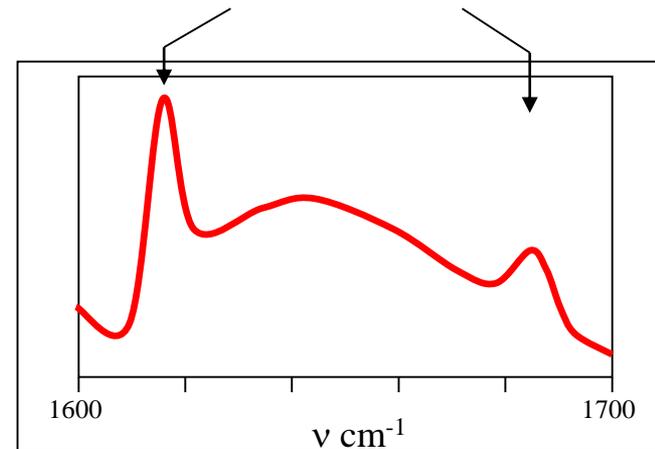
unordered

$\alpha$ -helix

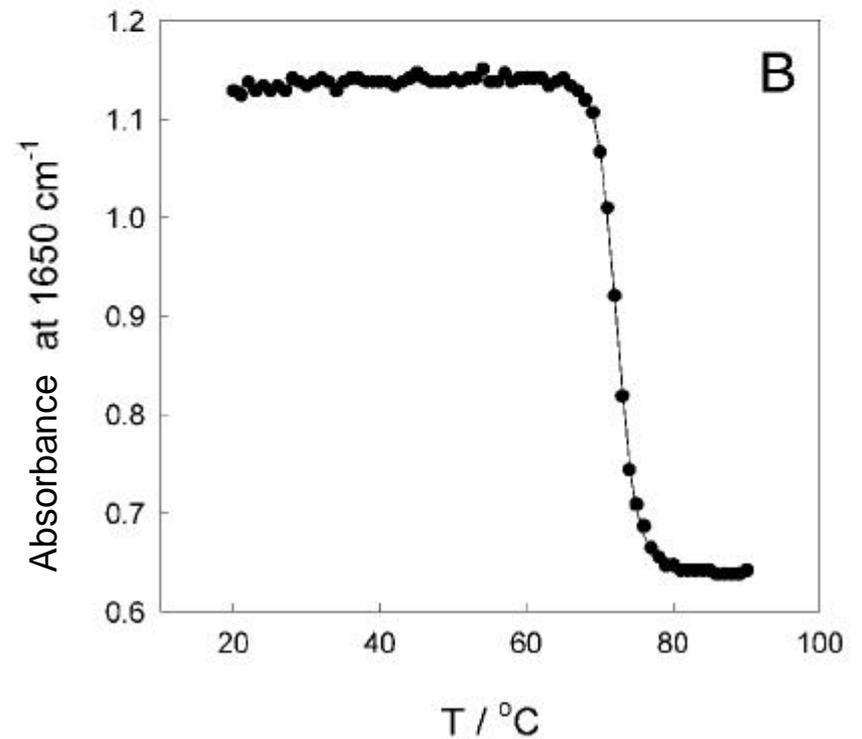
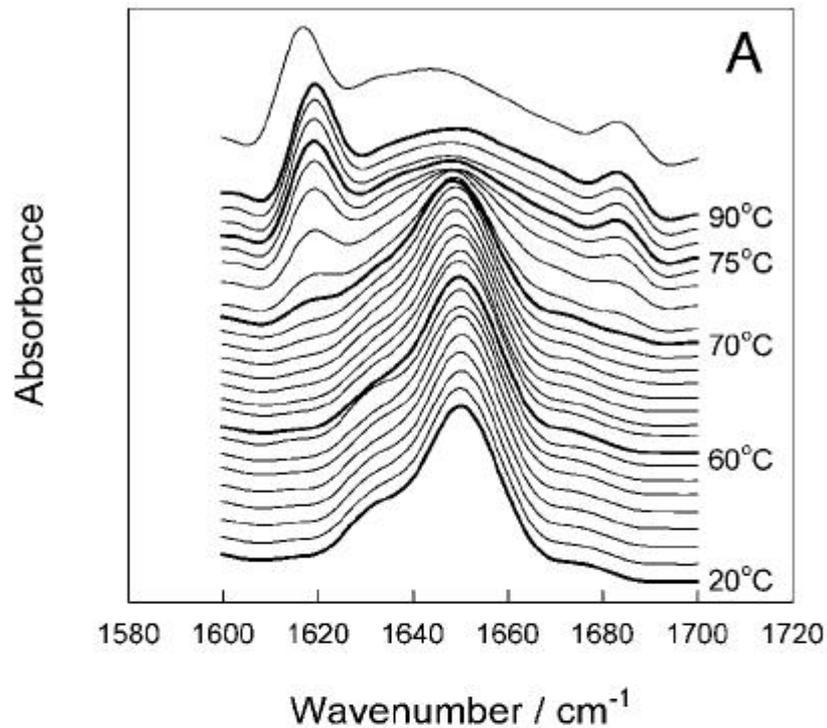


## Intermolecular interaction

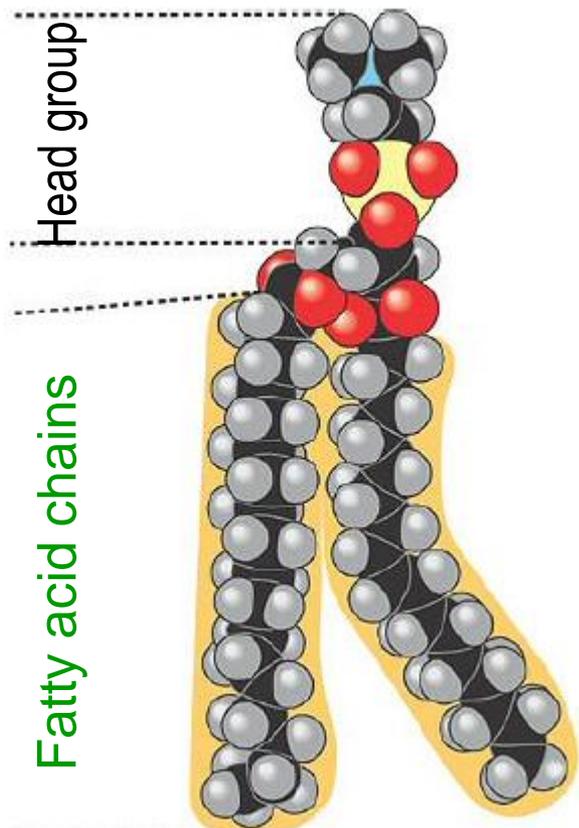
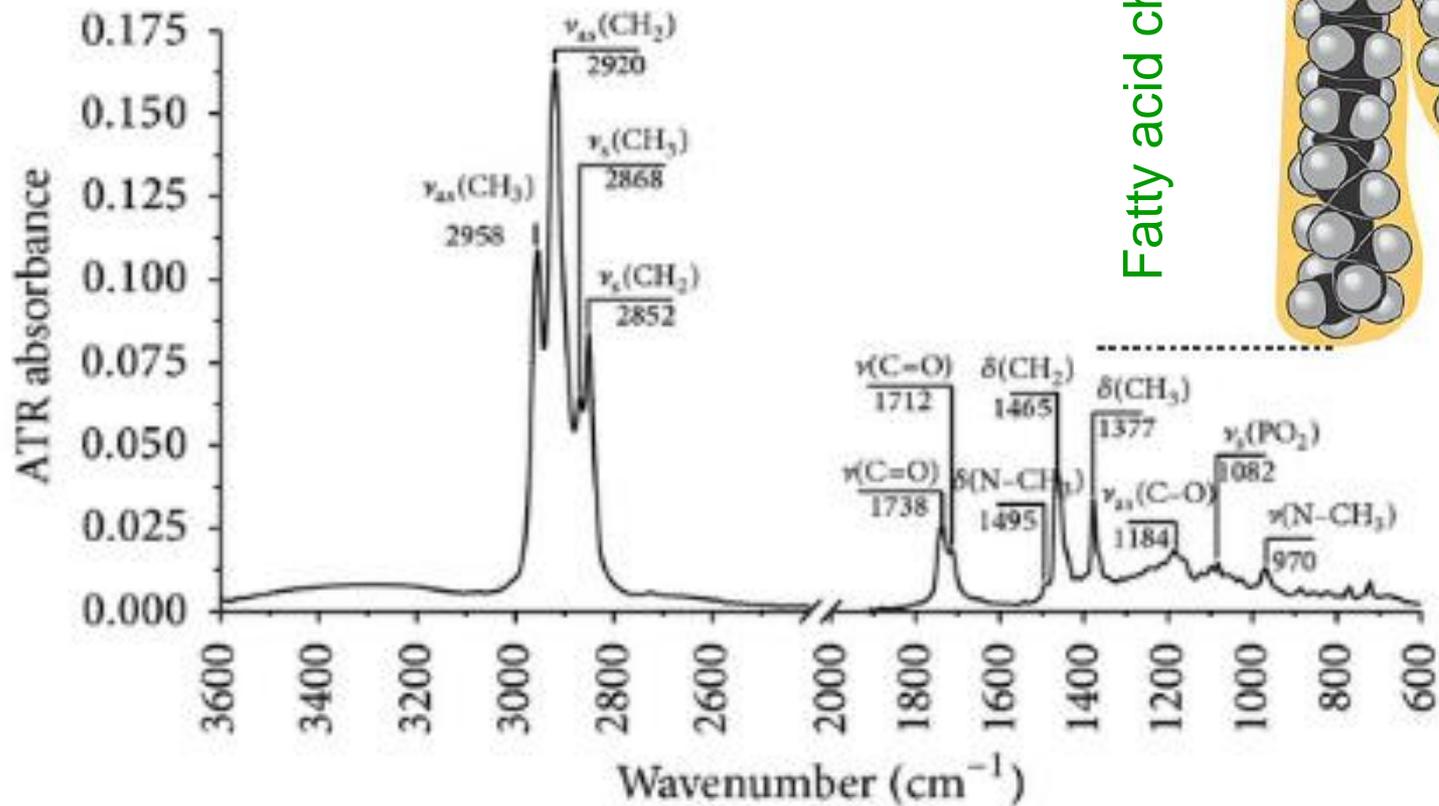
Intermolecular antiparallel  $\beta$ -sheet



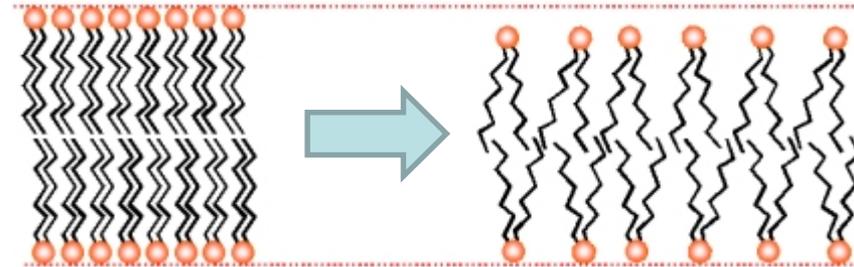
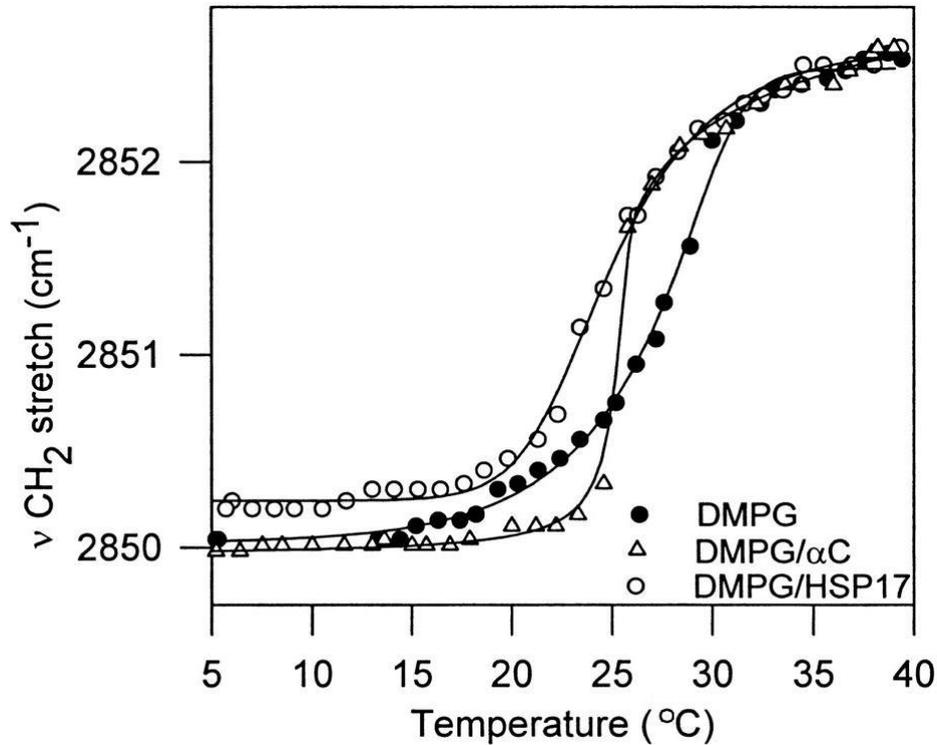
# Applications: Denaturation



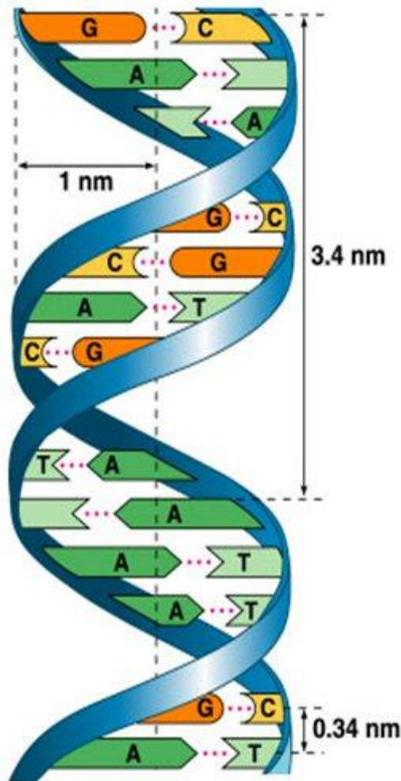
# Infrared spectroscopy of lipids



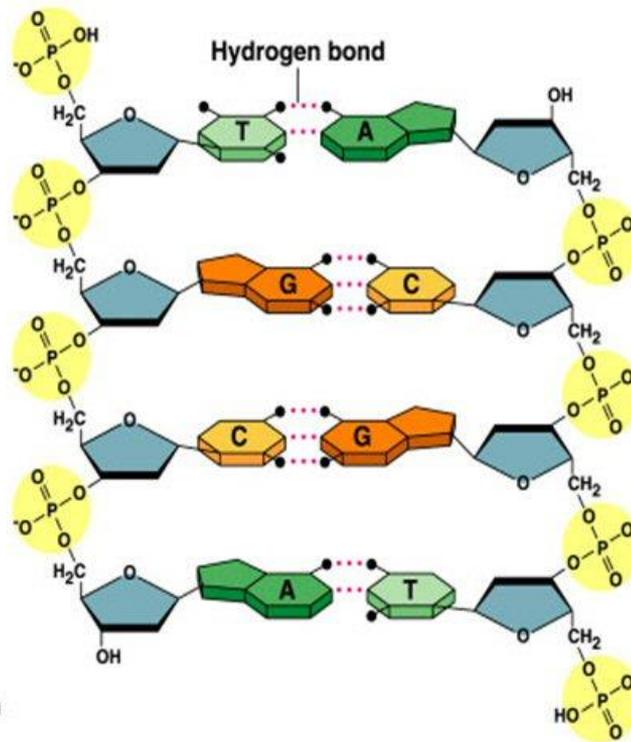
# Applications: lipid phase transitions



# Nucleic acids



(a) Key features of DNA structure



(b) Partial chemical structure

Bases (H bond)

Phosphate group.  
(PO<sub>2</sub><sup>-</sup>)

Sugar

# DNA

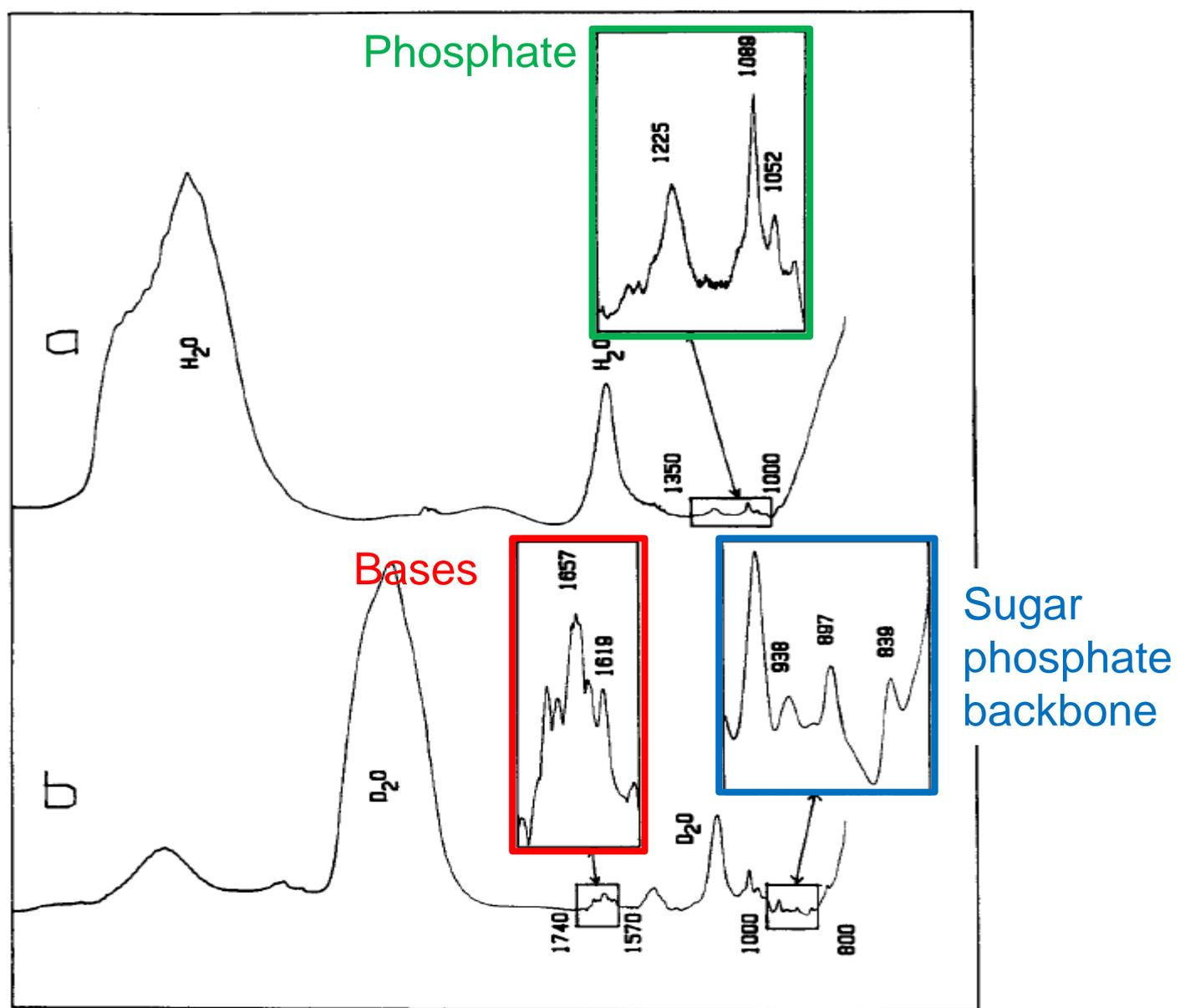


FIG. 1. FT-IR spectra of DNA in solution. (a) H<sub>2</sub>O solution; (b) D<sub>2</sub>O solution. Enlarged parts of the spectra present the absorptions involving mainly the vibrations of the phosphate groups (top), the double bonds of the bases in their plane (bottom left), and the sugar-phosphate backbone (bottom right).

# DNA, RNA A and B structures

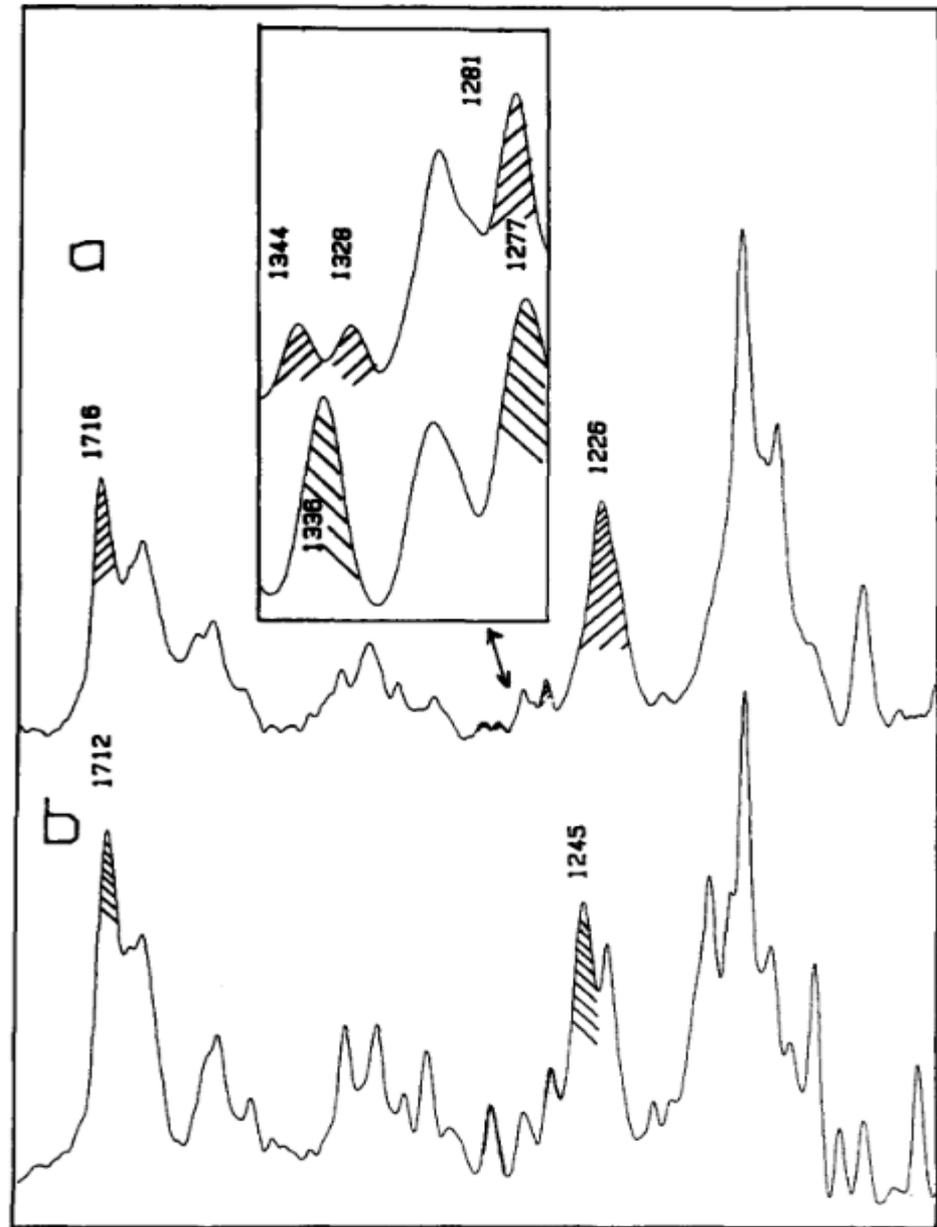
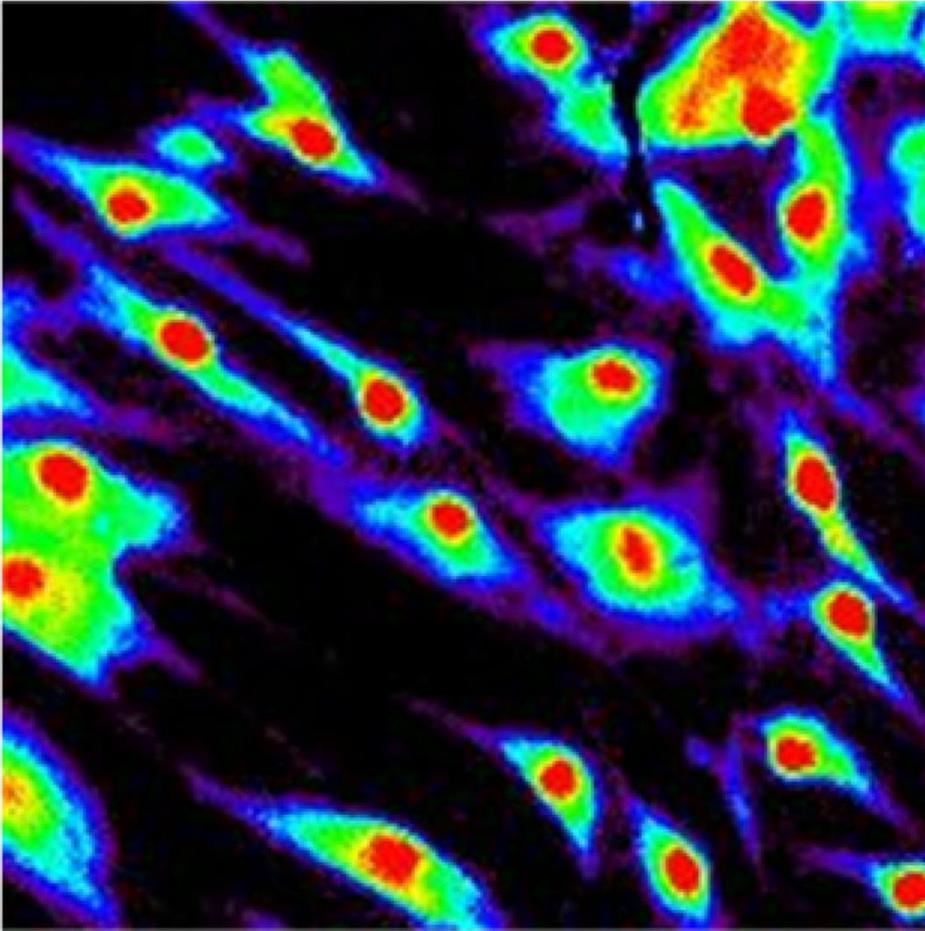


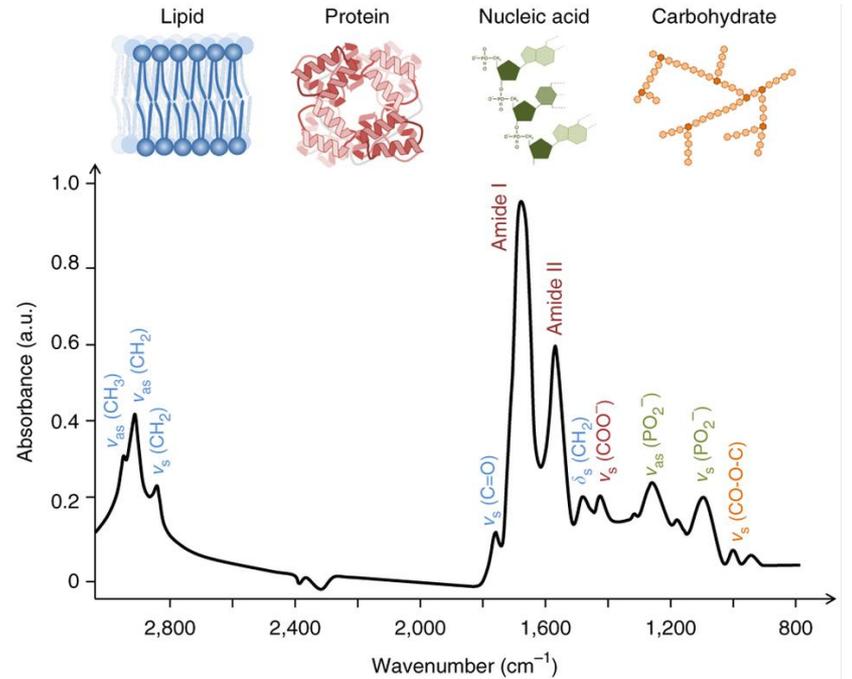
FIG. 3. FT-IR spectra of H<sub>2</sub>O solutions of d(A-U)<sub>n</sub> (a) and r(A-U)<sub>8</sub> (b). The enlarged area between 1350 and 1270 cm<sup>-1</sup> shows absorptions characteristic of A (\\) and B (//) geometries.

# Special IR methods: IR Microscope





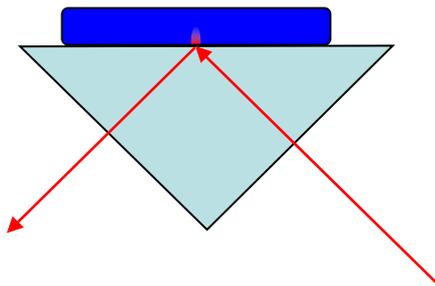
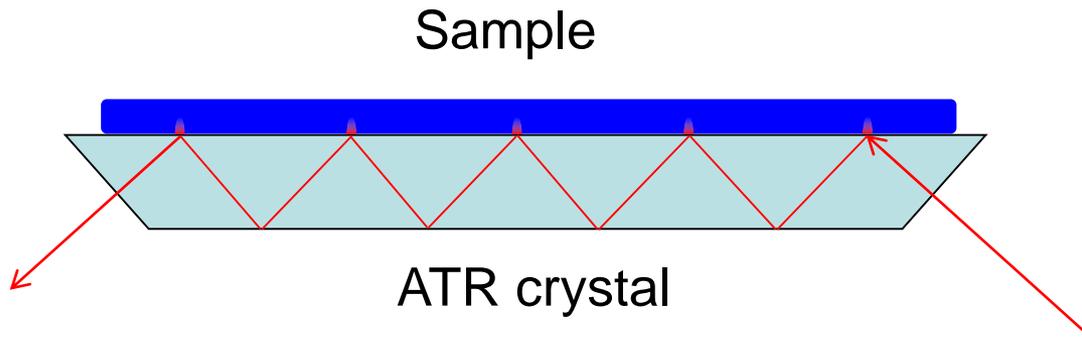
dermal fibroblasts imaged at  $1224\text{ cm}^{-1}$



# Portable FTIR spectrometer



# ATR technique (Attenuated Total Reflection)

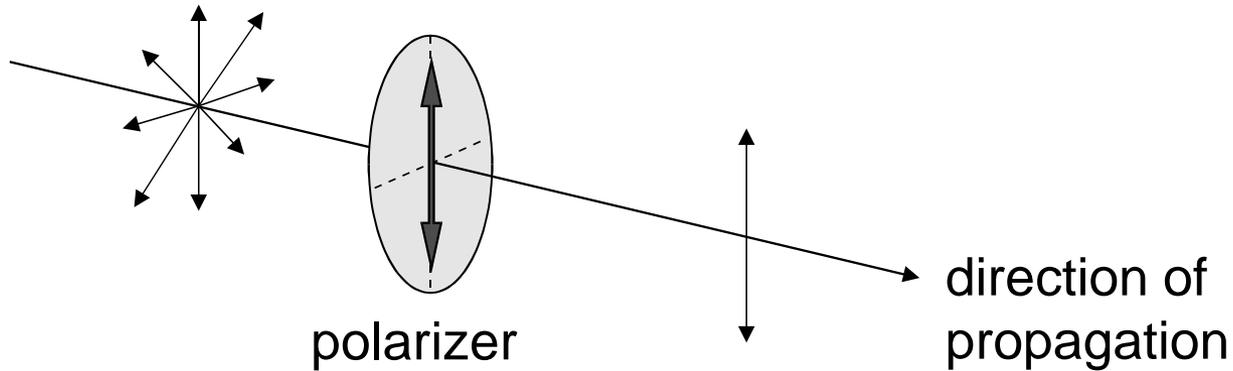


CD

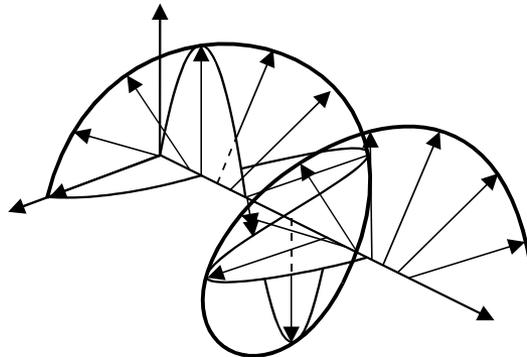
Circular dichroism spectroscopy

# Polarized light

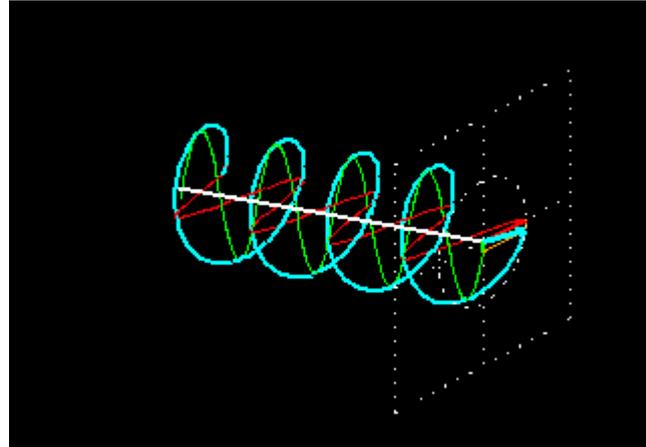
Linear polarized:



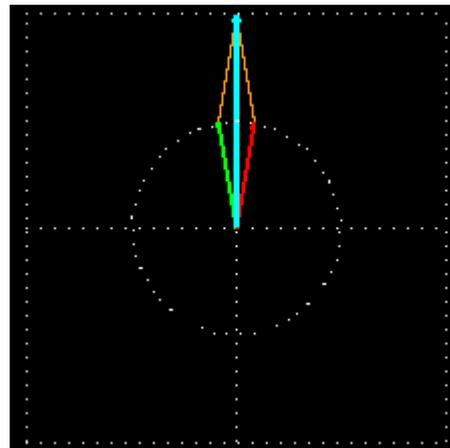
Circular polarized



Circular  
polarized light:



lin. pol light=  
right +  
left circ. pol.



Interaction of chiral molecules with left and right polarized light is different.

The absorbance difference is:

$$\Delta A = A_L - A_R = \Delta \epsilon \cdot c \cdot x$$

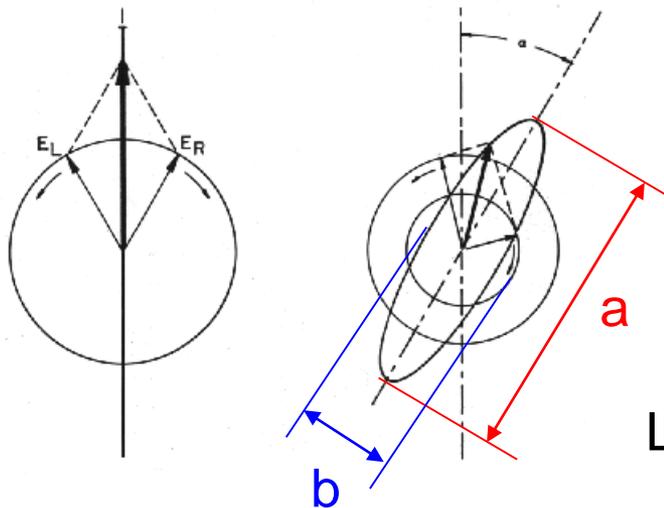
$$\Delta \epsilon = \epsilon_L - \epsilon_R$$

Ellipticity:  $\theta$      $\text{tg } \theta = b/a$

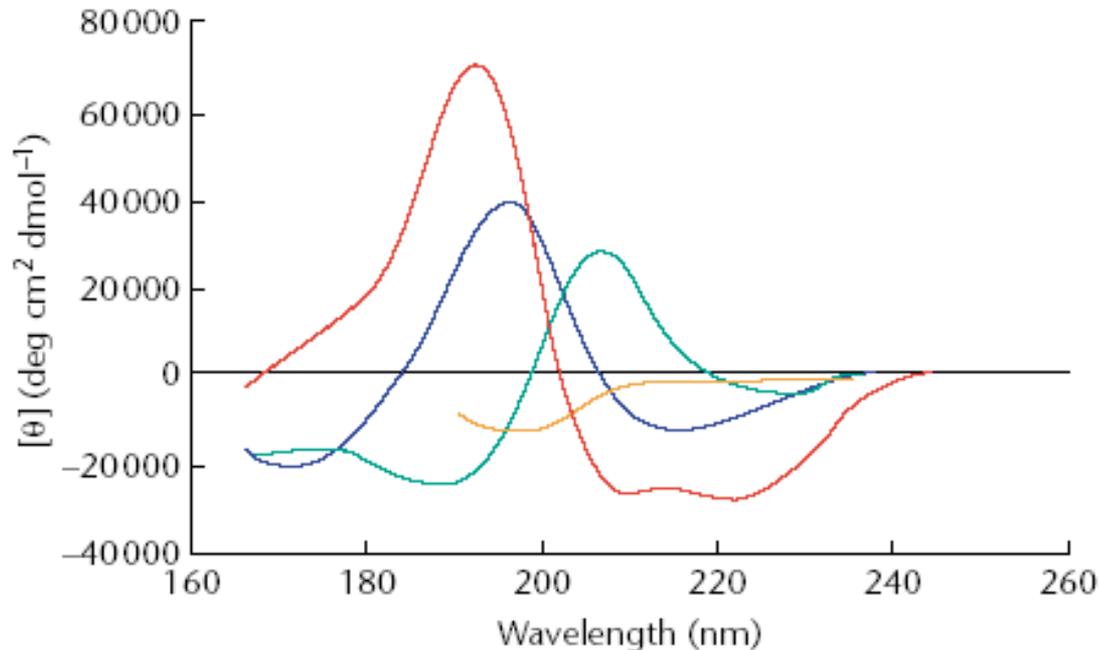
$$\theta = \frac{2.303}{4} \cdot (A_L - A_R) \cdot \frac{180}{\pi} \text{ [deg]}$$

Lambert-Beer like law:  $\theta = c \cdot l \cdot \theta_m$

( $\theta_m$ : molar ellipticity)



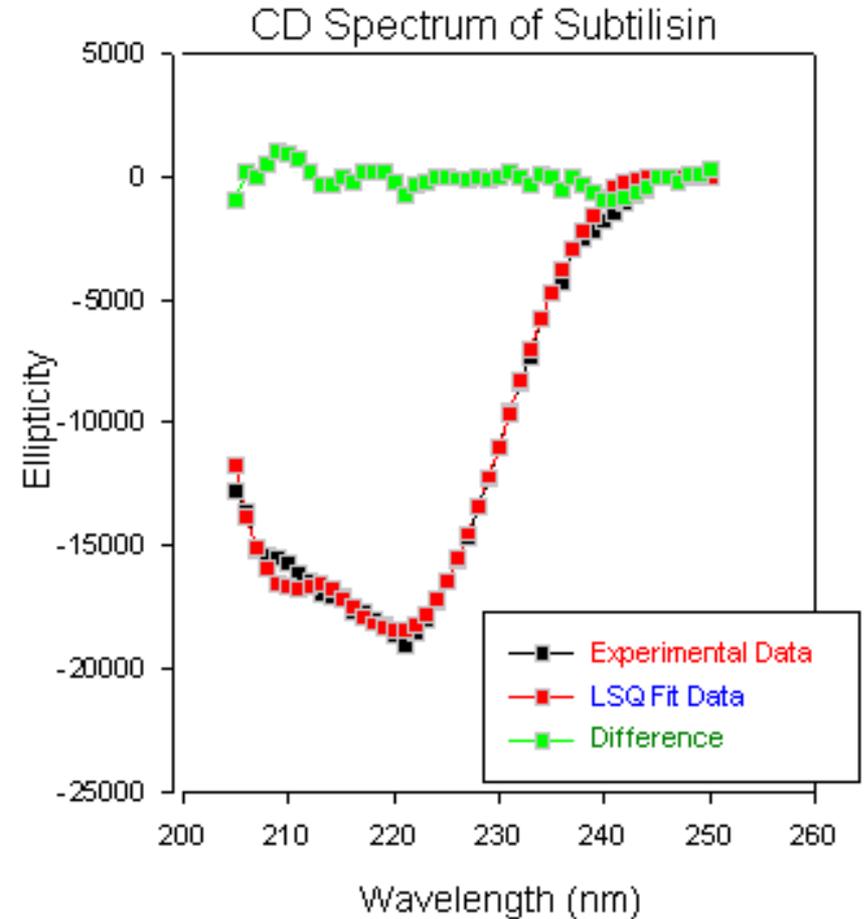
# CD and protein structure



The far-UV CD spectra associated with various types of secondary structure elements in proteins. **Red:  $\alpha$ -helix**; **blue: antiparallel  $\beta$ -sheet**; **green: type I  $\beta$ -turn**; **orange: irregular structure**.

(Data taken from the Encyclopedia of Life Sciences)

# The Structure and CD spectrum of Subtilisin



helix	sheet	coil
57.92	26.22	15.85

A) triosephosphate isomerase

B) hen egg lysozyme

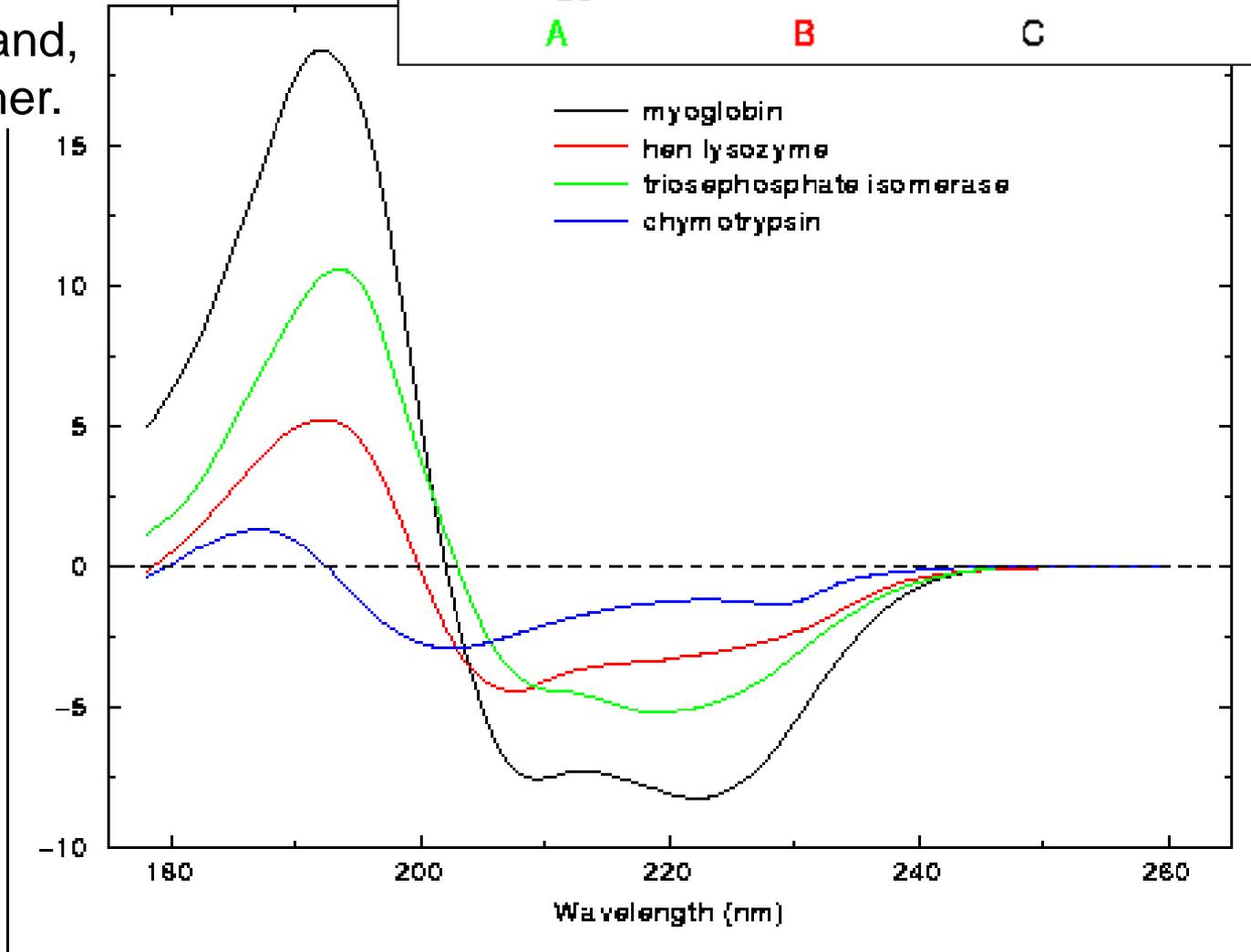
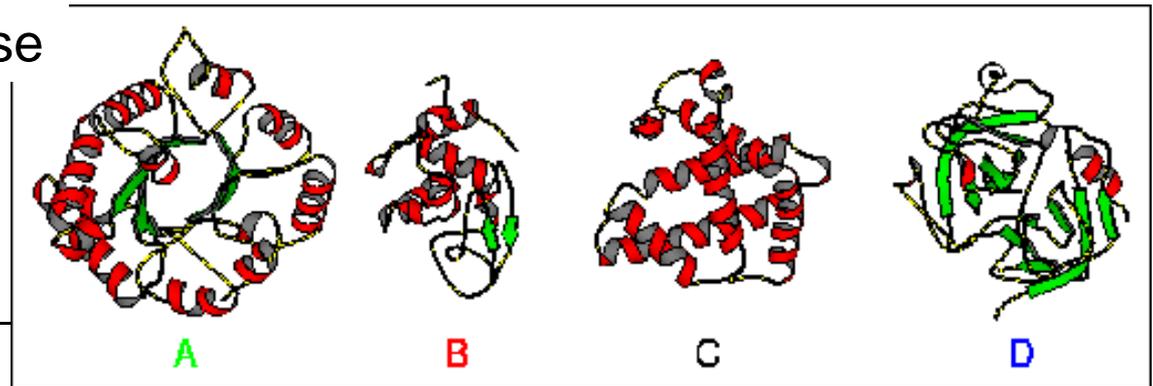
C) myoglobin

D) chymotrypsin

red: helix.

green: strand,

yellow: other.



End

# The CD spektrometer

