

**DESMIN PURIFICATION**

Based on Geisler, N. and Weber, K. Eur. J. Biochem. 111, 425-433, 1980.

**Number:****Date:****Buffers:**

<b>1. Buffer A</b>	2000 ml
40 mM imidazole-HCl (pH 6.9)	5.45 g
0.6 M KCl	89.46 g
1 mM EGTA	8 ml (0.25 M stock)
1 mM $\beta$ -mercaptoethanol (fresh)	140 $\mu$ l

to 500 ml, add 0.5 % Triton X-100 (2.5 ml)

<b>2. Extraction buffer</b>	1000 ml
10 mM imidazole-HCl (pH 6.9)	0.681 g
0.6 M KI	99.6 g
1 mM EGTA	4 ml (0.25 M stock)
0.5 % Triton X-100	5 ml
0.5 mM DTT (fresh)	0.5 ml (1 M stock)
0.2 mM ATP (fresh)	0.67 ml (300 mM stock)

<b>3. Urea Buffer</b>	1000 ml	2000 ml
6 M urea	360.36 g	720.72 g
10 mM Na-phosphate (pH 7.5)	1.42 g	2.84 g
5 mM EGTA stock)	20 ml	40 ml (0.25 M
0.1 % $\beta$ -mercaptoethanol	1 ml	2 ml

For salt gradient:

0.2 M KCl-Urea Buffer	100 ml
0.2 M KCl	1.49 g
Add Urea Buffer to 100 ml	

<b>4. Tris-acetate buffer</b>	1000 ml	2000 ml
0.01 M Trizma Base (pH 8.2 with acetic acid)	1.211 g	2.422 g
10 mM $\beta$ -mercaptoethanol	0.7 ml	1.4 ml

<b>5. Sodium-bicarbonate buffer</b>	1000 ml	2000 ml
1 mM NaHCO <sub>3</sub> (pH 8)	0.084 g	0.168 g
10 mM $\beta$ -mercaptoethanol	0.7 ml	1.4 ml

DEAE-cellulose (Whatman DE52): 16g swollen beads -&gt; 50 ml volume

For ELFO, put aside ~100  $\mu$ l samples, dissolve in ~5-  $\mu$ l 3x Laemmli solubilization buffer. If needed, add extra 1x Laemmli solubilization buffer (e.g., if there is substantial KDS precipitate).

**Steps:****Check:**

1. Excise chicken gizzard (~20 g)

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Weight of gizzard:

Set aside a small (~100 mg) piece; quick freeze in LN<sub>2</sub>, store at -80°C (for smitin ELFO)

2. Cut into small pieces

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Homogenize in 12.5 ml/g-tissue Buffer A (with Triton),

stir 30 min on ice

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4. Centrifuge, 9000 rpm, 10 min, GSA rotor

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(Alternatively, spin in Janetzky 3500 rpm)

5. Resuspend **pellet** in 12.5 ml/g-tissue "Buffer A without Triton"

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6. Stir 30 min on ice (ELFO #1)

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7. Centrifuge, 9000 rpm, 10 min, GSA rotor

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8. Repeat steps 5-7

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9. Resuspend **pellet** in 15 ml/g-tissue Extraction Buffer

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10. Stir for 6 hours, 4 °C

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11. Centrifuge, 9000 rpm, 10 min, GSA rotor

(Alternatively, Janetzky 3000 rpm, 4°C, 10 min)

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12. Repeat steps 9-11 2x (ELFO #2)

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13. Resuspend pellet in 3.75 ml/g-tissue Urea Buffer

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14. Stir at 37 °C for 2 hours to dissolve desmin

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15. Centrifuge, 9000 rpm, 10 min, GSA rotor

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16. Add 6-fold volume of ethanol to **supernatant**

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Supernatant volume (ELFO #3):

Ethanol volume:

17. Centrifuge, 9000 rpm, 10 min, GSA rotor

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17. Resuspend **pellet** in 1.25 ml/g-tissue Urea Buffer (ELFO #4)

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18. Chromatography: Sephadex G-25 (500 ml)

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Pump speed (Gilson): 600 (thin tubing)

AU Range: 0.1

Paper speed 1 mm/min

Approximate void volume: 600 ml

19. Chromatography: DEAE-cellulose (2.5 x 10 cm) (ELFO #5)

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Pump speed (Gilson): 400

paper speed 0.5 mm/min

AU range: 0.05

Elute with 200 ml salt gradient (0-0.2 M KCl in Urea buffer)

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Gradient mixer: 2 x 100 beakers connected with tubing.

20. Screen fractions for desmin (OD, SDS-PAGE)

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21. Pool desmin fractions (ELFO #6)

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22. Dialyze against Tris-acetate buffer

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23. Store (quick-freeze in LN<sub>2</sub>, store at -80 °C)

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